

Review

Environmental applications of ion chromatography

W. T. FRANKENBERGER, Jr.*

Department of Soil and Environmental Sciences, University of California, Riverside, CA 92521 (U.S.A.)

H. C. MEHRA

Chemical Waste Management, Inc., Kettleman City, CA 93239 (U.S.A.)

and

D. T. GJERDE

Sarasep, Inc., 1600 Wyatt Drive, Suite 10, Santa Clara, CA 95054 (U.S.A.)

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1. INTRODUCTION

Ion chromatography (IC) is a versatile, selective and sensitive method for the determination of a variety of anions and cations at trace and ultra-trace levels. It has been applied to hundreds of problems in various fields involving ionic analysis in clinical, food, pharmaceutical, industrial, plating solution and environmental samples. Perhaps the major reason for the rapid growth in popularity of IC is its usefulness in the environmental field. Due to the general complexity of environmental samples, IC has become an integral part in environmental analysis. Detection of specific trace elements at the sub-part-per-billion (10^9) level is a major challenge for the analytical chemist. The enhanced separating powers of the method also permits speciation of environmental contaminants. Different species of elements have different orders of toxicities.

Early publications (1970s) of IC work described environmental applications. Ion chromatography was introduced in 1975¹, but soon after two books were published in 1978 and 1979^{2,3}. There is a need to update and assemble new application papers in applied environmental sciences.

Several books^{4–8} and general review articles^{9–40} have been published on IC in recent years. The most recent book by Smith⁵ discusses IC applications up to 1986.

However, much of the work reviewed focuses on company information and lacks many of the published references. Several review articles on environmental applications have also been published⁴¹⁻⁵⁶.

Ion chromatography is a rapidly changing field. This review includes the most recent papers published in the literature as well as information available from the major IC instrument and column manufacturers on environmental pollution applications. The literature is reported according to applications in water, soil, sediment, sludge, plants, air (aerosols) and fossil fuels. A survey of the literature indicates that replication of the analysis can be a problem. Often good or unique work on specific elements can get "buried" with the description on separation of multiple ions. The speciation of different oxidation states of related ions and separation of less common ions have been emphasized in this review.

2. WATER ANALYSIS

IC has shown great promise for the sequential determination of ionic species in a wide variety of water samples. In many cases, little sample pretreatment is required. The technique has an intrinsic ability to cope with complex water samples with large quantities of matrix ions in the detection of the target solutes at trace or ultra-trace concentrations within the sample.

There are numerous reports on the use of IC in the determination of common anions such as Cl^- , NO_3^- , PO_4^{3-} and SO_4^{2-} in a wide range of water samples such as wastewater, rain water, drinking water, snow and Antarctic ice (Table 1). The use of high-performance liquid chromatography (HPLC) for the determination of organic anions and inorganic ions in such samples has been comprehensively reviewed by Bombaugh⁵⁷. The determination of organic and inorganic pollutants in different water samples using amperometric and colorimetric detection methods was recently reviewed⁵⁸. Amperometric detection tends to be selective and is very sensitive. The sensitivity and effectiveness of IC has also been compared to classical spectrometric methods⁵⁹. Many IC procedures have been standardized against colorimetric, titrimetric, steam distillation, autoanalyzer, atomic absorption spectrometry (AAS) and inductively coupled argon plasma emission spectrometry (ICAP).

In addition to separation and detection of various common anions, IC has been used for the determination of various uncommon species in water. Bromide has been selectively determined in water samples using amperometric detection⁶⁰. In a recent paper by Urasa and Nam⁶¹, chromium(III) and chromium(VI) in water samples were analyzed directly using cation- and anion-exchange columns employing a direct current plasma-AAS as an element-selective detector. Detectable concentrations of chromium species were in the low ppb range using preconcentration procedures. Cyanide, an extremely toxic ion which is used in electroplating and gold industries, has been determined in wastewater by IC using electrochemical detection^{41,107,110,114}. The simultaneous determination of CN^- , Cl^- , Br^- and I^- was recently reported with amperometric detection¹¹⁵.

Different oxidation states of some elements have different levels of toxicities and hence the separation and determination of these species is very important. While ICAP and AAS only detect the total elemental content, IC can determine different species simultaneously. IC has been useful for the speciation of toxic ions such as chromium^{61,124}, arsenic^{91,158} and selenium^{135,136}.

TABLE I
APPLICATIONS OF IC IN THE ANALYSIS OF WATER SAMPLES

R.S.D. = Relative standard deviation; Cond. = conductivity; GFAAS = graphite-furnace atomic absorption spectrometry; LOD = limit of detection; DAP = diaminopropionic acid; PDCA = pyridine dicarboxylic acid.

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Rain water, groundwater	Br ⁻	Dionex AS3	3.0 mM NaHCO ₃ ⁻ 2.0 mM Na ₂ CO ₃	Amperometric	LOD, 10 µg l ⁻¹ ; recovery, 97–110%; Cl ⁻ interferes if Cl ⁻ /Br ⁻ ratio > 1000:1	60
Rain, drinking water, lake and interstitial waters	NO ₃ ⁻ , SO ₄ ²⁻ , HCO ₃ ⁻ NO ₂ ⁻ , S ₂ O ₃ ²⁻ , Cl ⁻ Br ⁻ , H ₂ PO ₄ ⁻ , HCOO ⁻	Vydac 3021C	4.0 mM Sodium hydrogen phthalate	Cond.	Use of several different eluents	62
River water	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	TSK gel IC-anion PW	0.4 mM Trimellitate	UV-VIS	LOD, 0.1, 0.2 and 0.1 mg l ⁻¹ for Cl ⁻ , SO ₄ ²⁻ and NO ₃ ⁻ , respectively	63
Rain water	CH ₃ COO ⁻ , HCOO ⁻ , Cl ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex AS3	2.0 mM NaHCO ₃ ⁻ 1.67 mM Na ₂ CO ₃	Cond.	Detection of volatile organic acids and inorganic anions, simultaneously	64
Rain, well water, snow	NO ₂ ⁻	Shodex IC I-613	1.0 mM Phthalic acid-0.35 mM (C ₄ H ₉) ₄ NOH-0.5% tetrahydrofuran	Amperometric	LOD, 3 µg ml ⁻¹	65
Rain, wastewater, soil pore water	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ , total S	Vydac 302 IC	4.0 mM Phthalic acid (pH 5)	Cond.	LOD, 0.6, 0.2 and 0.2 µg ml ⁻¹ for Cl ⁻ , NO ₃ ⁻ and SO ₄ ²⁻ , respectively	66
Rain water, seawater	Dimethylsulfoxide	Bio-Rad HPX-87H	5.0 mM H ₃ PO ₄	UV (195 nm)	Ion exclusion; LOD, 10 µg l ⁻¹	67
Rain water, Antarctic ice	S(IV)	Dionex AS3	0.9 mM Bicarbonate-0.2% formaldehyde	Cond.	LOD, 0.1 µg l ⁻¹ , formaldehyde protects against oxidation	68

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TABLE 1 (*continued*)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Rain water	F ⁻ , Cl ⁻ , NO ₂ ⁻ , SO ₄ ²⁻ , Br ⁻ , NO ₃ ⁻ , PO ₄ ³⁻	Dionex anion exchange	3.0 mM NaHCO ₃ -2.4 mM Na ₂ CO ₃	Cond.	Semi-automatic (analysis of 60 samples per day)	69
River water	Arsenic anions	TSK gel IC-anion PW	NaNO ₂	Cond.	Recovery, 88–102%; LOD, 0.3–5 µg ml ⁻¹	70
Rain water	Na ⁺ , NH ₄ ⁺ , K ⁺	Dionex cation exchange	6.0 mM HNO ₃	Cond.	Significant variation between IC and AAS at lower ion concentrations	71
Simulated rain water	(i) F ⁻ , Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ , (ii) Na ⁺ , K ⁺ , NH ₄ ⁺ , Ca ²⁺ , Mg ²⁺	Dionex anion exchange (i) Dionex cation exchange	(i) 3.0 mM NaHCO ₃ -2.4 mM Na ₂ CO ₃ (ii) 5.0 mM HCl	(i) Cond. (ii) Cond.	Required 1–4 ml sample within 0.05–20 mg l ⁻¹ range	72
Rain water	Na ⁺ , K ⁺ , NH ₄ ⁺ , Ca ²⁺ , Mg ²⁺	Dionex	NaHCO ₃ -Na ₂ CO ₃	Cond.	Step-gradient elution; lacks sensitivity for Mg ²⁺	73
River water	HCO ₃ ⁻ , CO ₃ ²⁻	TSK gel IC-Anion PW	0.4 mM Trimel-litate (pH 7.5)	UV (270 nm)	Compared well with Floco injection analysis	74
Rain, lake	(i) SO ₄ ²⁻ , Cl ⁻ , NO ₃ ⁻ , F ⁻ , PO ₄ ³⁻ , (ii) Li ⁺ , Na ⁺ , K ⁺ , NH ₄ ⁺ , Ca ²⁺ , Mg ²⁺	Dionex AS4 HPIC-CS3	(i) 1.8 mM Na ₂ CO ₃ -2.2 mM NaHCO ₃ (ii) 4.8 mM HCl-4.0 mM DAP·HCl-4.0 mM histidine	(i) Cond. (ii) Cond.	Cost-effective compared to spectroscopy methods; LOD, 0.005–0.05 mg l ⁻¹	75
Rain water	Alkali and alkaline earth cations	Dionex	35 mM HCl	Cond.	LOD, 0.05–0.15 µmol l ⁻¹ for alkali ions and 0.2–0.6 µmol l ⁻¹ for alkaline earth ions	76
Rain water	Na ⁺ , Cl ⁻	Bio-Rad S-X2 and Dionex AS4	Lithium hydrogen phthalate-phthalic acid	Cond.	Simultaneous determination of anions and cations; LOD, R.S.D. <0.5%	77
Natural and wastewater	NO ₃ ⁻	C ₁₈ reversed phase	60 mM KH ₂ PO ₄ -15 mM H ₃ PO ₄	UV (210 nm)	NO ₂ ⁻ and organic chthoniophores do not interfere; LOD, 7 µg l ⁻¹	78

River and drinking water	Cl^- , NO_3^-	Spherisorb ODS-2 (Phase separations, Queensferry, U.K.)	Sodium hydrogen phthalate	UV	High pH and high ionic strength 79 of eluent not recommended; LOD, 3–4 nmol
River water	NO_3^- , NO_2^-	Dionex AS4	4.0 mM Na_2CO_3^- 4.0 mM NaHCO_3	UV (220 nm)	Time of analysis for NO_3^- , 5 min 80
Lake, sea, well and tap waters	F^-	Shodex IC I-524A	2.5 mM phthalic acid (pH 4.0)	Cond.	Microdiffusion of F^- from sample matrix, LOD, $4 \mu\text{g l}^{-1}$
River, drinking water	Li^+	HPLC-CS2	8.0 mM HCl	Cond.	LOD, $1 \mu\text{g l}^{-1}$ 82
River water	Na^+ , K^+	Zorbax SHL	0.01 M Lithium acetate	Cond.	Time of analysis, 10 min 83
Pond water	Ca^{2+} , Mg^{2+} , $(\text{Sr}^{2+}$, Mn^{2+} , Fe^{2+} , Co^{2+} , Ni^{2+} , Zn^{2+} , Cd^{2+} , Cu^{3+})	Dionex cation exchange	1.0 mM $\text{Ba}(\text{NO}_3)_2$	Cond.	Al^{3+} or Cr^{3+} did not elute under the conditions described 84
River water	Ca^{2+} , Mg^{2+}	Synthesized polystyrene–divinylbenzene	0.12 M HClO_4	VIS (590 nm)	Post-column reactor 85
Pond water	Ca^{2+} , Mg^{2+}	Bio-Rad AG1-X8	1.0 mM $\text{Pb}(\text{NO}_3)_2$	Cond.	Good agreement with AAS 86
River water	Al^{3+}	Dionex CS2	0.2 M $(\text{NH}_4)_2\text{SO}_4$ (pH 2.8)	UV (310 nm)	Post-column derivatization; LOD, $7 \mu\text{g l}^{-1}$ 87
Tap water	Al^{3+}	Dionex CS2	0.1 M K_2SO_4 (pH 3.0)	Fluorescence	Post-column reaction; LOD, $1 \mu\text{g l}^{-1}$ 88
River and sewage waters	NH_4^+	Hitachi 2632 (anion-exchange resin)	Water	Coulometric	Results agree with phenate method; LOD, ng levels 89
River water	Quaternary ammonium compounds	Whatman Partisil PAC 10 (cyano–amino bonded phase)	CHCl_3 – CH_3OH	Cond.	LOD, $0.02 \mu\text{g l}^{-1}$ ($50 \mu\text{l}$ sample size) 90
River water	AsO_3^{3-} , AsO_2^{4-} , SeO_3^{2-} , SeO_4^{2-}	C_{18} reversed phase	1.0 mM Hexadecyltri-methylammonium bromide	GFAAS	LOD, 15–25 ng 91

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TABLE I (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Seawater	F ⁻ , Cl ⁻ , Br ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex anion exchange	2.5 mM Na ₂ CO ₃ ⁻ 3.0 mM NaHCO ₃	Cond.	R.S.D. 0.4–4.2%	92
Seawater	Ni ²⁺ , Co ²⁺ , Cu ²⁺	Waters Nova-Pak C ₁₈	10 mM CH ₃ COONa–CH ₃ OH–H ₂ O (76:24%) (pH 8.4)	UV (340 nm)	Time of analysis, 20 min	93
Seawater	I ⁻	TSK gel IC-anion PW (polymethacrylate, anion-exchange capacity, 0.03 meq)	0.1 M NaCl + 5 mM Sodium phosphate buffer (pH 6.7)	Amperometric	LOD, 5 µg l ⁻¹ ; R.S.D. 3%	94
Seawater	Ca ²⁺ , Mg ²⁺ , Sr ²⁺ , Ba ²⁺	Waters IC-Pak-C	10 mM Phenyl-ethylamine (pH 5.5)	Cond. and UV/VIS	LOD, 0.2–21.0 ng ml ⁻¹ ; aromatic base eluents were assessed	95
Surface water	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Waters IC-Pak-A	Borate–gluconate	Cond.	System peaks were avoided by treating sample with cation exchanger	96
Surface water	NO ₃ ⁻ , NO ₂ ⁻	Merck LiChrosphere-RP ₁₈	n-Octylamine (pH 6)	UV	Good agreement with photometric determination	97
Groundwater	Se(IV) and Se(VI)	Dionex AS3	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	Preservation of sample techniques discussed	98
Pore water	Cl ⁻ , F ⁻ , NO ₃ ⁻ , Br ⁻ , SO ₄ ²⁻	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 2.0 mM Na ₂ CO ₃	Cond.	Results compared with colorimetric methods; LOD, 0.005–0.1 mg l ⁻¹ ; recovery, 97–103%	99
Natural water	F ⁻ , Cl ⁻ , Br ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Waters IC-Pak-A	Borate–gluconate, KOH and lithium benzoate	UV/VIS or Cond.	Time of analysis, 16 min	100

Antarctic snow and ice	NO_3^- , SO_4^{2-} , Cl^- , Na^+ , NH_4^+ , K^+	Dionex anion exchange; Dionex cation exchange	2.0 mM Na_2CO_3^- 2.5 mM NaHCO_3 (anions) 5.0 mM HCl (cations)	Cond.	Efforts were made to reduce contamination problems	101
Antarctic ice	(i) Cl^- , HCOO^- , methane sulfonate, NO_2^- , NO_3^- , SO_4^{2-} (ii) CH_3COO^- , F^- , Na^+ , NH_4^+ Cl^- , NO_3^-	(i) Dionex 030-985 anion exchange (iii) Wescan 269-004 cation exchange	(i) 0.7 mM NaHCO_3^- 2.0 mM Na_2CO_3 (ii) 4.0 mM HNO ₃	(i) Cond. (ii) Cond.	Sample preservation info. Time of analysis, 30 min	102
Wastewater	NO_3^- , Cl^- , SO_4^{2-} , PO_4^{3-}	HKS-1	1.0 mM Tyrosine (pH 10.8)	Cond.	LOD, 10 ng ml ⁻¹ ; R.S.D. < 2%	103
River water	NO_3^- , Cl^- , SO_4^{2-} , PO_4^{3-}	Dionex AS1	36 mM NaHCO_3^- 24 mM Na_2CO_3	Cond.	Complex Fe with cyanide to avoid hydroxide interference	104
Wastewater	Cl^- , NO_3^- , PO_4^{3-} , SO_4^{2-}	Dionex 30827	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	Compared with various other methods	105
Wastewater	Cl^- , SO_4^{2-} , PO_4^{3-} , $\text{S}_2\text{O}_3^{2-}$, SCN^- , NO_2^- , NO_3^-	Dionex AS3 or AS5	2.55 mM NaHCO_3^- 2.05 mM Na_2CO_3	Cond.	Time of analysis, 15 min	106
Wastewater	CN^-	Dionex AS4	2.2 mM Na_2CO_3	Cond.	Oxidation of CN^- to CNO^- by hypochlorite	107
Wastewater	SO_3^{2-} , SO_4^{2-} , $\text{S}_2\text{O}_3^{2-}$	Bio-Rad Bio-gel TSK IC-anion PW	Gluconate-borate (pH 7.6)	UV (254 nm)	Compares with conventional conductivity and electrochemical detection	108
Wastewater	F^- , Cl^- , NO_3^- , SO_4^{2-} , PO_4^{3-}	Dionex 30170	2.1 mM NaHCO_3^- 1.7 mM Na_2CO_3	Cond.	Na_2CO_3 fusion used to decompose sample; LOD, 8–10 μg	109
Wastewater	CN^-	Dionex AS3	14.7 mM Ethylene-diamine-10.0 mM Na_2HBO_3 –1.0 mM Na_2CO_3	Amperometric	LOD, 0.02 mg l ⁻¹ ; compares well with colorimetric determination	110
Saline water	NO_3^- , PO_4^{3-}	Dionex anion exchange	3.0 mM NaHCO_3^- 1.9 mM Na_2CO_3	Cond.	LOD, 0.1 mg l ⁻¹	111
Natural water	S^{2-}	Dionex-AS3	H_3BO_3 –NaOH–ethylene-diamine	Amperometric	LOD, 1.9 ng ml ⁻¹ ; R.S.D. 0.64%	112

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TABLE I (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Natural water	F^- , Cl^- , NO_3^- , SO_4^{2-}	Anion-exchange resin	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	R.S.D. 1.3%; LOD, 30, 40, 240, 400 pg for F^- , Cl^- , NO_3^- and SO_4^{2-} , respectively	113
Wastewater	CN^- (Cl^- , ClO_3^- , SO_4^{2-})	Dionex anion exchange	2.2 mM Na_2CO_3	Cond.	Free CN^- and metal CN^- complexes determined	114
Lake water	CN^- , Cl^- , Br^- , I^-	Vydac 302 IC	Potassium hydrogen phthalate (5.0 mM, pH 4.3)	Amperometric	Simultaneous determination; LOD, 0.2–12 $\mu\text{g l}^{-1}$; recovery 95–105%; time of analysis, 10 min	115
Wastewater	PO_4^{3-}	Dionex anion exchange	3.0 mM Na_2CO_3^- 1.0 mM NaOH	Cond.	LOD, 1.5 mg l^{-1} ; R.S.D. 0.49%	116
Refinery wastewater	NH_4^+ , ethanolamines	Dionex MPIC NS-1	5.0 mM hexane- sulfonic acid	Cond.	Time of analysis, <11 min; R.S.D. 2%; LOD, 0.05–5 mg l^{-1}	41
Wastewater	(i) SCN^- , $\text{S}_2\text{O}_3^{2-}$, (ii) $\text{S}_4\text{O}_6^{2-}$, SO_3^{2-} , SO_4^{2-}	Dionex anion exchange	(i) 2.4 mM Na_2CO_3^- 3.0 mM NaHCO_3 (ii) 7.2 mM Na_2CO_3^- 9.0 mM NaHCO_3	Cond.	Long elution time; badly tailing peaks for $\text{S}_2\text{O}_3^{2-}$; recovery, 97–101%; R.S.D. 3.8%	117
Wastewater	Borate	Dionex anion exchange	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	R.S.D. 0.05–2.6%; recovery, 96.4–99.3%; LOD, 0.05 mg l^{-1}	118
Wastewater	F^- , Cl^- , PO_4^{3-} , NO_3^- , SO_4^{2-}	Dionex anion exchange	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	Time of analysis, 24 min	51
Wastewater	F^- , Cl^- , NO_3^- , HPO_4^{2-} , SO_4^{2-} , I^- , SCN^- , ClO_4^-	Biotronik IC-1000	1.0 mM Tyrosine	Cond.	R.S.D. <0.02%; LOD, 0.01 $\mu\text{g ml}^{-1}$	119
Natural and wastewater	F^- , Cl^- , NO_2^- , PO_4^{3-} , Mg^{2+} , Ca^{2+} , HCO_3^- , CO_3^{2-}	Dionex IC16	Na_2CO_3 or HNO_3	Cond.	Linear response	120

Wastewater	NH ₄ ⁺	Dionex CS1	H ₃ BO ₃	Fluorescence	121
Wastewater	SO ₄ ²⁻ (F ⁻ , Cl ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , PO ₄ ²⁻ , and Br ⁻)	Dionex anion exchange	NaHCO ₃ -Na ₂ CO ₃	Cond.	122
Wastewater	Cu ²⁺ , Pb ²⁺ , Zn ²⁺ , Ni ²⁺ , Co ²⁺ , Cd ²⁺ , Fe ²⁺ , Mn ²⁺	Waters μBondapak C ₁₈	2.0 mM NaOS-30 mM tartaric acid-20 mM citric acid (pH 3.4)	VIS (520 nm)	123
Wastewater	Cr(VI) + Cr(III)	Dionex CS5	2.0 mM PDCA, 2.0 mM Na ₂ HPO ₄ , 10 mM NaI, 50 mM CH ₃ CO ₂ NH ₄ , 2.8 mM LiOH	VIS (520 nm)	124
Acid rain	F ⁻ , Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex AS3	NaHCO ₃ -Na ₂ CO ₃	Cond.	125
Drinking water	NO ₃ ⁻ , (F ⁻ , Cl ⁻ , PO ₄ ³⁻ , SO ₄ ²⁻ , Br ⁻ , NO ₂)	Dionex AS1	4.5 mM NaHCO ₃ -2.4 mM Na ₂ CO ₃	Cond.	126
Natural water	F ⁻ , Cl ⁻ , PO ₄ ³⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex AS1	3.0 mM NaHCO ₃ -2.4 mM Na ₂ CO ₃	Cond.	59
Natural water	F ⁻ , Cl ⁻ , NO ₂ ⁻ , H _x PO ₄ ^(3-x) , SO ₄ ²⁻	PRPX-100 poly-styrene-divinylbenzene	0.8 mM Phthalate (pH 6.8)	UV (265 nm)	129
Drinking water	NO ₂ ⁻	High capacity ion-exchange resin	4.0 M NaCl	UV (210 nm)	128
Drinking water, rain, lake water	NO ₂ ⁻	Anion exclusion/HS	5.0 mM H ₂ SO ₄	Amperometric	127
Drinking water	(i) NO ₃ ⁻ (ii) NO ₂ ⁻	Waters IC-Pak Dionex HPIC AS4	5.0 mM LiOH 28 mM NaHCO ₃ -22 mM Na ₂ CO ₃	UV (214 nm) Cond.	130

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TABLE I (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Natural water	F ⁻ , Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex anion exchange	3.0 mM NaHCO ₃ -2.4 mM Na ₂ CO ₃	Cond.	R.S.D. <2%	131
Drinking and pond water	NO ₃ ⁻	Partisil 10 SAX	1.0 mM Potassium hydrogen phthalate	UV (265 nm)	LOD, 0.1 µg ml ⁻¹	132
Natural water	NO ₃ ⁻ , SO ₄ ²⁻	Dionex anion exchange	0.5 mM Potassium hydrogen phthalate	Cond.	Time of analysis, 20 min includes cleanup procedure to remove humic substances	133
Tap water	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Toyo Soda IC anion PW	2.0 mM Tiron	Cond. or UV (290 nm)	R.S.D. 2%; time of analysis, 10 min	134
River and drinking water	Se(IV), Se(VI)	Dionex anion exchange	2.5 mM Na ₂ CO ₃ ⁻ 2.0 mM KOH	Cond.	Also detects TeO ₄ ²⁻ , TeO ₃ ²⁻ , PO ₄ ³⁻ , NO ₃ ⁻ , SO ₄ ²⁻ ; time of analysis, 30 min; R.S.D. <0.7%	135
River and drinking water	Se(IV), Se(VI)	Nucleosil 55B (Gasuburo Kogyo, Tokyo)	75 mM Ammonium phosphate	Fluorescence	Post-column derivatization; time of analysis, 10 min; LOD of Se(IV), 0.17 µg l ⁻¹	136
Tap water	F ⁻	Interaction ORH-301	0.001 M H ₂ SO ₄	Cond.	Ion exclusion; time of analysis, <15 min	137
Drinking water	HCO ₃ ⁻	Interaction Ion-310	Deionized water	Cond.	Anion-exclusion method; time of analysis, <10 min	137
Drinking water	F ⁻ , Cl ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , Br ⁻ , PO ₄ ³⁻ , SO ₄ ²⁻	Dionex anion exchange	2.0 mM Na ₂ CO ₃ ⁻ 0.75 mM NaHCO ₃	Cond.	Recovery, 90-108%; R.S.D. 1-6%	138
Drinking water	Br ⁻ , I ⁻ , NO ₂ ⁻ , SCN ⁻	Nucleosil 10-CN	Cetyltrimethylammonium bromide	Amperometric	Ion pair C ₁₈ ; LOD for I ⁻ or SCN ⁻ , 1 µg l ⁻¹	139
Natural water	MoO ₄ ²⁻ , WO ₄ ²⁻	Dionex anion exchange	6.0 mM Na ₂ CO ₃	Cond.	LOD, 1 µg l ⁻¹ ; R.S.D. 15%; time of analysis, 17 min	140
Tap water	Al ³⁺	Dionex CG2	0.1 M K ₂ SO ₄ (pH 3.0)	Fluorescence	Post-column derivatization; LOD, 1 µg l ⁻¹ ; R.S.D. 3.4%; time of analysis, 2 min	141

Drinking water	(i) NH_4^+ , Ca^{2+} , Mg^{2+} (ii) Ca^{2+} , Mg^{2+} , Sr^{2+} , Ba^{2+}	(i) Vyddac SC cation exchanger (ii) Resin BN-X4 blend	(i) 1.25 mM HNO_3 (ii) 1.0 mM Ethylene diamine dinitrate (pH 6.1)	(i) Cond. (ii) Cond.	(i) Time of analysis, 10 min (ii) Time of analysis, 5 min; R.S.D. < 2.5%
Drinking water	Ca^{2+} , Mg^{2+} (Ni^{2+} , Zn^{2+} , Pb^{2+} , Mn^{2+} , Sr^{2+})	Dionex CS-2	2.0 mM Ethylenediamine-2.0 mM citric acid (pH 4.0)	Cond.	LOD, < 20 $\mu\text{g l}^{-1}$
Drinking water	(i) Cl^- , NO_3^- , SO_4^{2-} (ii) Ca^{2+} , Mg^{2+}	(i) Waters IC-pak™ anion (ii) Cation columns	(i) 2.0 mM Ethylenediamine-octane-sulfonic acid (pH 6) (ii) 3.0 mM Sodium octanesulfonate	(i) Cond. (ii) Cond.	Simultaneous analysis of anions and cations; time of analysis, 16 min
Drinking water	(i) Na^+ , K^+ , NH_4^+ (ii) Mg^{2+} , Ca^{2+}	(i) Cation exchanger (ii) Cation exchanger	(i) HNO_3 (ii) Ethylenediamine	LOD, 1 mg l^{-1}	145
Drinking water	NO_3^- , NO_2^- Cr(III) and Cr(VI)	Waters μ Bondapak C ₁₈ Dionex-CS2 and Dionex-AS7	5.0 mM Tetramethyl ammonium phosphate 7.5 mM Trilithium citrate-10 mM oxalic acid	UV (214 nm) Direct current plasma-AAS	Time of analysis, 6 min; LOD, 0.1 mg l^{-1}
Drinking water Natural and industrial process stream samples	NO_3^- , SO_4^{2-}	Dionex AS1	2.4 mM Na_2CO_3^- 3.0 mM NaHCO_3	Cond.	Also detected F^- , Cl^- , Br^- , PO_4^{3-} ; R.S.D. 3%
Fog samples	(i) Cl^- , NO_3^- , NO_2^- , SO_3^{2-} , SO_4^{2-} (ii) F^- , CH_3COO^- , HCOO^- (iii) Na^+ , NH_4^+ , K^+ (iv) Ca^{2+} , Mg^{2+}	(i) Dionex AS4 (ii) Dionex AS4 (iii) Sykam LCA K01 (iv) Sykam LCA K01	0.5 mM NaHCO_3^- 1.3 mM Na_2CO_3 1.5 mM $\text{Na}_2\text{B}_4\text{O}_7$ 4.5 mM HNO_3 1.0 mM Histidine-1.0 mM diaminopropionic acid-12 mM HCl	(i) Cond. (ii) Cond. (iii) Cond. (iv) Cond.	LOD, < 20 $\mu\text{g l}^{-1}$, time of analysis, 12 min Time of analysis, 10 min Time of analysis, 7 min Time of analysis, 10 min

(Continued on p. 222)

TABLE 1 (*continued*)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Antarctic ice	(i) CH_3SO_3^- , HCOO^- , F^- , Cl^- , NO_3^- , SO_4^{2-}	(i) Dionex AS4	(i) 0.65 mM NaHCO_3^- , 2.5 mM NaHCO_3^- , 4.0 mM Na_2CO_3	(i) Cond.	Time of analysis, 6 min; requires 5 ml sample to reach $10^{-10} \text{ g g}^{-1}$	149
	(ii) Na^+ , NH_4^+ , K^+	(ii) Dionex CS2	(ii) 25 mM HCl	(ii) Cond.		149
Wastewater	Organic acids	Dionex AS2	0.01 M HCl	Cond.	Ion exclusion	149
	Fe^{3+} , Cu^{2+} , Zn^{2+} , Co^{2+} , Fe^{2+} , Mn^{2+} , alkaline earth cations	Silica-based cation exchanger	0.1 M Tartrate (pH 3.05)	VIS	Post-column derivatization; LOD, $2 \mu\text{g l}^{-1}$	150
Demineralized water	CN^-	Dionex AS3	1.0 mM Na_2CO_3^- –10 mM $\text{NaH}_2\text{BO}_3^-$ –15 mM ethylenediamine	Amperometric	Application to photographic wastes	151
	CN^- , S^{2-} , Br^-	Dionex AS3	1.0 mM Na_2CO_3^- –10 mM $\text{NaH}_2\text{BO}_3^-$ –15 mM ethylenediamine	Amperometric (Ag electrode)	R.S.D. <2%; time of analysis, 8 min	41
Wastewater	Cd^{2+} , Co^{2+} , Mn^{2+}	Dionex CS5	3.0 mM Pyridine-2,6-dicarboxylic acid–4.3 mM LiOH	UV (520 nm)	LOD, $0.1 \mu\text{g l}^{-1}$; recovery, 97–114%; R.S.D. 0.57%	152
	Organic acids (formic, acetic, propionic, oxalic, malonic, succinic glutaric acid)	Dionex AG3, AS3, AF3	0.1 M Octane sulfonic acid	Cond.	R.S.D. 3–4.9%; LOD, $4\text{--}10 \text{ ng ml}^{-1}$	153
Rain water	F^- , CH_3COO^- , HCOO^-	Dionex AX1	2.0 mM $\text{Na}_2\text{B}_4\text{O}_7$	Cond.	Time of analysis, 15 min; R.S.D. 3.7–7.1%; LOD, $0.05\text{--}0.91 \text{ mg l}^{-1}$	154
	NO_3^-	Biotronik BT II AN	$2 \cdot 10^{-3} \text{ M } \text{Na}_2\text{CO}_3$	UV (215 nm)	R.S.D. 0.1%; LOD, 5 ng g^{-1}	155
Snow samples (Antarctic)						

Seawater (artificial)	I ⁻	TSK gel IC anion SW separator	0.5 mM Sodium citrate (pH 5.0)	UV/Cond.	R.S.D. 4%; LOD, 3 ng; also detected Cl ⁻ , NO ₃ ⁻ , ClO ₄ ⁻ ; Cl ⁻ , Br ⁻ , NO ₃ ⁻ may cause interference if present in large quantities	156
Natural water	I ⁻	SAX-I	30 mM Na ₂ CO ₃	UV (226 nm)/ Cond.	Also separates Cl ⁻ , PO ₄ ³⁻ , Br ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ ; LOD, 0.005 µg (100 µl); recovery, 95%	157
Natural water	As(III) + As(V)	Bio-Rad organic analysis column (Annex HPX-87H ion exclusion)	0.01 M Orthophosphoric acid	Amperometric/ UV (200 nm)	R.S.D. 1.3%; detection was linear up to 1.0 µM; time of analysis, 20 min	158
River water	Na ⁺ , K ⁺	Zorbex SIL	0.01 M Lithium acetate	Cond.	LOD, 0.005 M; time of analysis, 10 min	159
Purified water	F ⁻ , Cl ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , Br ⁻ , PO ₄ ³⁻ , SO ₄ ²⁻	Dionex AS4	2.0 mM Na ₂ CO ₃ ⁻ 0.75 mM NaHCO ₃	Cond.	LOD, 3–10 µg l ⁻¹ ; recovery, 90–107%; R.S.D. 0.4–6.3%	160
Lake water	NO ₃ ⁻	Dionex AS2	Water	UV (210 nm)	R.S.D. 0.45%; time of analysis, 10 min; LOD, low ppb	161
Natural water	Cl ⁻ , Br ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Wescan anion exchange (269-001)	Sodium hydrogen phthalate	Cond.	LOD, 1 mg l ⁻¹ ; chromatography on a guard cartridge	162
Wastewater effluent	PO ₄ ³⁻	Dionex AS4	2.25 mM NaHCO ₃ ⁻ 2.0 mM Na ₂ CO ₃	Cond.	Other anions separated Cl ⁻ , F ⁻ , NO ₂ ⁻ , Br ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	163

Organic acids appear in wastewater as a result of biological decomposition of organic materials. Ion-exclusion chromatography was found to be extremely reliable in the determination of weak organic acids in various water samples^{102,153}. McDowell and Stedman⁴⁴ reported the interference of carbonic acid in their studies on the determination of formic, acetic, propionic and butyric acids in various water samples. Ethanolamines have also been determined using IC⁴³.

Another serious pollution problem is acid rain¹²⁵. IC has been applied in the analysis of acid rain to determine the anion and cation composition. There are also several reports on the ionic analysis of Antarctic snow samples^{101,102}. Sensitivity is an extremely important parameter since the ion content in these samples can be about 100 times lower than that of comparable samples taken from industrial areas of the northern hemisphere⁵.

3. SOIL SEDIMENT, SLUDGE AND PLANT ANALYSIS

IC analysis of the ionic species in complex systems such as soil, sediments, sludges and plants has received little attention in recent years. Soil, sediment and sludge materials are often analyzed with the objective to determine a single constituent. However, the composition of other ions present may be determined by IC. The advantages of IC in the analysis of such complex samples over conventional chemical techniques include the determination of a variety of ions in a single run, speed and speciation and minimal sample preparation. Table 2 shows the environmental applications of IC in soil, sediment, sludge and plant samples.

Dick and Tabatabai¹⁷⁰ were the first to determine NO_3^- and SO_4^{2-} in soil samples using suppressed IC after extraction with LiCl. Inorganic nitrogen and sulfur are of prime importance in soils since they are essential plant nutrients. Frankenberger and co-workers have used IC for the determination of the following ions in soil: Cl^- , NO_2^- , NO_3^- , PO_4^{3-} , SO_3^{2-} , SO_4^{2-} , CN^- , Br^- , I^- , Li^+ , Na^+ , NH_4^+ , K^+ , Ca^{2+} , Mg^{2+} , Ba^{2+} (refs. 115, 167, 168, 185) as well as Sr^{2+} (ref. 185), SeO_3^{2-} (refs. 174, 175), SeO_4^{2-} (refs. 173, 175) AsO_4^{3-} (ref. 177), WO_4^{2-} , (ref. 183), MoO_4^{2-} (ref. 182), CrO_4^{2-} ref. 180) H_2BO_3^- (ref. 190) and HCO_3^- (ref. 190). Recently, Mehra and Frankenberger¹⁷⁵ developed an IC method to separate and determine SeO_3^{2-} and SeO_4^{2-} , simultaneously in soils. Speciation of Se is important in understanding its potential toxicity and mobility in groundwater and soils. Simultaneous determination of arsenite and arsenate has also been reported in sediments with no interferences from other anions¹⁹⁶.

IC is becoming an attractive alternative to the conventional spectroscopic methods for determination of metal ions in soil. Meaney *et al.*²⁰⁵ used IC for the determination of Fe^{3+} and Al^{3+} in soil and clay samples. Bertsch and Anderson¹⁸⁸, using a fully automated IC system, determined Al^{3+} extracts using a UV-visible detector. Alkali and alkaline earth cations in soil have been detected using conductometric detection both by single column and suppressed techniques. Reversed-phase columns have been used for determination of Pb, Zn, Co, Fe and Mn in soils¹²³. Determination of total Cu, Ni and Zn in soils has recently been reported by (i) digestion of soil with HF, HClO_4 and HNO_3 followed by (ii) extraction of metals as dithiozionate in CHCl_3 , (iii) destruction of the metal dithiozinate complex with HNO_3 and (iv) final determination by IC using a chromogenic reagent for the detection of the metals by visible photometry¹⁸⁷.

TABLE 2
APPLICATIONS OF IC IN THE ANALYSIS OF SOIL, SEDIMENT, SLUDGE AND PLANT SAMPLES

R_s = Resolution. MS = mass spectrometry; DAN = 2,3-diaminonaphthalene.

Sample	Tion	Analytical column	Eluent	Detector	Comments	Ref.
Soil	SO_4^{2-} (total S)	Dionex AS4	18 mM NaHCO_3^- 1.2 mM Na_2CO_3^-	Cond.	Time of analysis, 10 min; recovery, 84–98%; sample prep., Na_2O_2 fusion	164
Soil	SO_4^{2-}	(i) Waters IC-Pak A (ii) Vydac 302 IC	(i) Borate-gluconate buffer (ii) 4 mM Phthalic acid (pH 4.5)	(i) Cond. (ii) Cond.	Compared extractants: R.S.D. 1.9–8.4%; compared to ICAP; recovery, 97–108%	165
Soil	NO_3^-	Dionex AS4	0.75 mM NaHCO_3^- 2.2 mM Na_2CO_3^-	Cond.	LOD, 0.1 ng N dm^{-3} ; Cl^- interference eliminated with Ag^+ cation-exchange resin; compared well with steam distillation	166
Soil	PO_4^{3-}	Vydac 302 IC	1.5 mM Phthalic acid (pH 2.7)	Cond.	LOD, 0.3 $\mu\text{g l}^{-1}$; time of analysis, 20 min; R.S.D. 1.1% (500 μl); compared well with autoanalyzer	167
Soil	Cl^- , NO_3^{2-} , SO_4^{2-} (NO_2^- , SO_3^{2-})	Vydac 301 IC	4.0 mM Phthalic acid	Cond.	Time of analysis, 11 min; LOD, 0.025–1.0 ng l^{-1} ; R.S.D. 3.6–8.4%; compared well to conventional methods	168
Soil	Cl^- , NO_3^- , SO_4^{2-}	Wescan Ion-guard anion cartridge (269-003)	15 mM Phthalic acid	UV (300 nm)	Time of analysis, 3 min	169
Soil	NO_3^- , SO_4^{2-}	Dionex anion exchange	3.0 mM NaHCO_3^- 1.8 mM Na_2CO_3^-	Cond.	Time of analysis, 12 min; R.S.D. 0.4–4.3%; compared well with methylene blue	170

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TABLE 2 (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Soil, plants	Cl ⁻ , NO ₃ ⁻ , PO ₄ ³⁻ , SO ₄ ²⁻	Apex ODS	0.5 mM Tetrabutyl ammonium hydroxide, 50% methanol, potassium hydrogen phthalate	UV (255 nm)	(SO ₄ ²⁻) and steam distillation (NO ₃ ⁻); recovery, 98–102%; LOD, 0.2 µg ml ⁻¹	171
Soil, plants	NO ₃ ⁻ (NO ₂ ⁻)	Dionex AS4	16 mM Na ₂ CO ₃ ⁻ 16 mM NaHCO ₃	UV (220 nm)	Recovery, 84–108%; time analysis, 20 min; interference by aspartic acid	172
Soil	Se (total)	Cosmosil 5SL	Cyclohexane–ethyl acetate (95:5)	Fluorescence	Time of analysis, 5 min	130
Soil	SeO ₄ ²⁻ (NO ₃ ⁻ , NO ₂ ⁻ , SO ₄ ²⁻)	Vydate 302 IC	4.0 mM Phthalic acid (pH 4.6)	Cond.	Post-column derivatization (DAN); time of analysis, 6 min.; R.S.D. 3.9%; recovery, 93–97%	173
Soil	SeO ₃ ²⁻ (Cl ⁻ , NO ₃ ⁻ , NO ₂ ⁻ , PO ₄ ³⁻)	Vydate 302 IC	1.5 mM Phthalic acid (pH 2.7)	Cond.	Applicable with high SO ₄ ²⁻ levels; R.S.D. 1.46 (500 µl); LOD, 18 µg l ⁻¹ ; compared well with AAS and ICAP	174
Soil	SeO ₃ ²⁻ and SeO ₄ ²⁻ (Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻)	Wescan resin-based (269-029)	4.0 mM p-Hydroxybenzoic acid	Cond.	Applicable with high Cl ⁻ levels; R.S.D. 2% (500 µl); LOD, 3 µg l ⁻¹ ; time of analysis, 8 min; comparable to AAS and ICAP	175
Soil	F ⁻ , Cl ⁻ (NO ₂ ⁻ , PO ₄ ³⁻ , NO ₃ ⁻ , SO ₄ ²⁻)	Dionex AS3	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	Humic materials removed through soil phase extraction; LOD, 60–110 µg l ⁻¹ ; R.S.D. 0.9–1.86% (500 µl); time of analysis, 15 min	176

Soil	AsO_4^{3-} (Cl^- , NO_3^- , PO_4^{3-} , SO_4^{2-})	Wescan resin-based (269-029)	6.0 mM <i>p</i> -Hydroxybenzoic acid	Cond.	AsO_3^{3-} was not detected; R_s between AsO_4^{2-} and PO_4^{2-} 1.93; LOD, 92 $\mu\text{g l}^{-1}$; time of analysis, 20 min; compared well with AAS and ICAP	177
Soil	Br^- , SCN^- , I^-	Whatman Partisil 10 SAX	5.0 mM KH_2PO_4^- 10% CH_3CN organic modifier	UV (205 nm)	Interference with NO_3^- ; time of analysis, 23 min; LOD, 1–3 ng	178
Soil	CN^-	Waters IC-Pak anion	5.0 mM KOH	Amperometric	LOD, 100 ng ml^{-1}	179
Soil	CN^- , Cl^- , Br^- , I^-	Vydac 302 IC	Potassium hydrogen phthalate (5.0 mM, pH 4.3)	Amperometric	Simultaneous determination; LOD, 0.2–12 $\mu\text{g l}^{-1}$; recovery 95–105%; time of analysis, 10 min	115
Soils, sludges	CrO_4^{2-} (NO_3^- , SO_4^{2-})	Wescan resin-based (269-029)	5.0 mM <i>p</i> -Hydroxybenzoic acid (pH 8.5)	Cond.	Organic impurities removed through solid phase extraction; R.S.D. 1.98% (500 μl); LOD, 92 $\mu\text{g l}^{-1}$, compared well with ICAP; time of analysis, 16 min	180
Soil	Cl^- , NO_3^- , HCO_3^- , SO_4^{2-}	Vydac anion exchange	3.0 mM Potassium hydrogen phthalate	Cond.	Samples included soil, and drinking water.	181
Soil	MoO_4^{2-} (Cl^- , NO_3^- , PO_4^{3-} and SO_4^{2-})	Waters TSK gel IC-Pak Anion 1 (26770)	5.0 mM <i>p</i> -Hydroxybenzoic acid (pH 8.25)	Cond.	LOD, 45 $\mu\text{g l}^{-1}$; time of analysis, 15 min; R.S.D. 2.7% (500 μl)	182
Soil, sludge	WO_4^{2-} (Cl^- , NO_3^- , SO_4^{2-})	Wescan resin-based (269-029)	5.0 mM <i>p</i> -Hydroxybenzoic acid (pH 8.5)	Cond.	LOD, 170 $\mu\text{g l}^{-1}$; time of analysis, 13 min; R.S.D. 1.9% (500 μl); R_s , WO_4^{2-} and SO_4^{2-} , 2.84	183
Soil	Li^+ , Na^+ , NH_4^+ , K^+	DuPont ZIPAX SCX	3.0 mM HNO_3	Cond.	Li^+ / Na^+ separation poor; time of analysis, 7 min; LOD, 0.1–5.0 mg l^{-1}	184
Soil	Li^+ , Na^+ , NH_4^+ , K^+	Vydac 401 TP cation exchange	1.0 mM HNO_3 (pH 2.1)	Cond.	LOD, 0.05–1 mg l^{-1} ; time of analysis, 5 min; compared well with conventional methods	185

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TABLE 2 (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Soil	Mg ²⁺ , Ca ²⁺ (Sr ²⁺ , Ba ²⁺)	Vydac 401 TP cation exchange	5.0 mM Ethylenediamine dinitrate (pH 6.1)	Cond.	LOD, 0.1–1 mg l ⁻¹ ; time of analysis, 4 min; compared well with conventional methods	186
Soil	Ca ²⁺ ; Mg ²⁺ (Sr ²⁺ , Mn ²⁺ and Zn ²⁺)	Dionex CS2	2.0 mM Ethylenediamine–2.0 mM citric acid	Cond.	LOD for Ca ²⁺ and Mg ²⁺ were 143 20 and 10 µg l ⁻¹ , respectively	143
Soil	Ca ²⁺ , Mg ²⁺ (Sr ²⁺ , Mn ²⁺ , Fe ²⁺ , Ni ²⁺ , Cu ²⁺ , Zn ²⁺ , Cd ²⁺)	Dionex cation exchange	0.3 mM Ba(NO ₃) ₂	Cond.	Time of analysis, 10 min; compared well with AAS; LOD, 10 ⁻⁶ M	84
Soil	Li ⁺ , Na ⁺ , K ⁺ , Rb ⁺ , Cs ⁺	Dionex cation exchange	5.0 mM HCl	Cond.	Time of analysis, 12 min; compared well with AAS and flame photometry; R.S.D. 2.3–6.9%	186
Soil	Mg ²⁺ , Ca ²⁺ , Sr ²⁺ , Ba ²⁺	Dionex cation exchange	2.5 mM HCl–2.5 mM m-phenylenediamine dihydrochloride	Cond.	Time of analysis, 16 min; compared well with AAS; R.S.D. 2.3–6.9%	186
Soil	Cu ²⁺ , Ni ²⁺ , Zn ²⁺	Dionex CS5	4.0 mM Pyridine 2,6-dicarboxylic acid–50 mM CH ₃ COOH–CH ₃ COONa (pH 4.8)	UV (520 nm)	Detection as dithizone complexes; compared well with AAS	187
Soil	Al ³⁺	Dionex CG3	0.4 M NH ₄ Cl (pH 3.1)	UV-VIS	Post-column reaction; LOD, 20 µg l ⁻¹ (100 µl); time of analysis, 3 min; compared well with AAS	188
Soil	Pb, Zn, Co, Fe, Mn	Waters µBondapack C ₁₈	2.0 mM NaOS–10 mM tartaric acid (pH 3.4)	UV (520 nm)	Post-column reaction; time of analysis, 20 min	123
Soil	Alkyl methyl-phosphonic acid	Dionex MPIC-NSI	2.0 mM Tetrabutylammonium hydroxide–1.0 mM Na ₂ CO ₃ –10–24% CH ₃ CN	Cond.	Time of analysis, 15 min; LOD, 0.2 mg kg ⁻¹	189

Soil and sediment	Borate, bicarbonate	Wescan ion exclusion column	D-Sorbitol	Cond.	Time of analysis, 6 min; R.S.D. 0.66–1.42%; LOD for borate, $0.1 \mu\text{g l}^{-1}$	190
Soil	F^- , Cl^- , NO_2^- , PO_4^{3-} , Ca^{2+} , Mg^{2+}	Dionex anion-exchange resin	Na_2CO_3 – NaHCO_3	Cond.	Linear response was reported	120
Soils, plants	MoO_4^{2-} (WO_4^{3-})	Cosmosil C ₁₈	1.5 mM Tiron–30 mM tetrabutylammonium bromide	UV (315 nm)	Time of analysis, 11 min; recovery, 97–105%; R.S.D. 2.4–4.5%	191
Plants	NO_3^-	Wescan anion exchange	4.0 mM Phthalic acid	Cond.	Used for determination of ¹⁵ N	192
Plants	NO_3^- , SO_4^{2-} (Cl^- , PO_4^{3-} , Br^-)	Hamilton PRP X-100	0.5 mM Pyromellitate buffer (pH 3)	UV (295 nm)	LOD, 2–5 ng; time of analysis, 10 min	193
Plants	Cl^- , NO_3^- , PO_4^{3-} , SO_4^{2-}	Dionex AS4	2.0 mM Na_2CO_3 0.7 mM NaHCO_3 in 2% 2-propanol	Cond.	Time of analysis, 10 min	194
Plants	(i) Li^+ , Na^+ , NH_4^+ , K^+ Rb^+ , Cs^+ , Ca^{2+} , Mg^{2+} (ii) Ca^{2+} , Mg^{2+}	(i) Dionex CS3 (ii) Dionex CS3	30 mM HCl–1.0 mM 2,3-diaminopropionic acid–0.5 mM ZnCl_2	(i) Cond. (ii) Cond.	Time of analysis, 8 min	194
Plants			27.5 M HCl–2.25 mM L-histidine · HCl		Time of analysis, 12 min	194
Plants	Cl^- , PO_4^{3-} , NO_3^- , SO_4^{2-} , glycolate, malate	Dionex AS3	2.8 mM NaHCO_3 2.2 mM Na_2CO_3	Cond.	Time of analysis, 12 min; R.S.D. 0.1–2.1%; compared well to NO_3^- electrode	195
Plants	Cl^- , HPO_4^{2-} , NO_3^- , SO_4^{2-} , malate, oxalate	Dionex AS4	2.8 mM NaHCO_3 2.2 mM Na_2CO_3	Cond.	R.S.D. <5%; recovery, 95–104%; compared well with conventional methods; time of analysis, 16 min; sample: tobacco	196
Plants	(i) K^+ , Na^+	(i) Dionex cation exchange	(i) 5.0 mM HCl	(i) Cond.	Time of analysis, 8 min; compared well with AAS and photometry; R.S.D. 0–7.69%	197

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TABLE 2 (continued)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Plants	(ii) Ca^{2+} , Mg^{2+}	(ii) Dionex cation exchange	(ii) 2.5 mM HCl -2.5 mM <i>m</i> -phenyldiamine dihydrochloride	Cond.	Time of analysis, 8 min; compared well with AAS; R.S.D. 0.7-14%	197
	HCOOH	Alltech Model OA-1000 organic acid column (58°C)	5 mM H_2SO_4	UV (214 nm)	Time of analysis, 12 min; similar results with GC-MS	198
Sediments	Cl^- , NO_3^- , SO_4^{2-}	Dionex AS4	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	High Cl^- concentration interferes with NO_3^- and SO_4^{2-} determination; time of analysis, 25 min	199
Sediments	I ⁻	Dionex AS3	20 mM NaNO_3^- 25 mM NaOH	Amperometric (Ag^+ electrode)	R.S.D. 3.5%; recovery 90-111%; time of analysis, 10 min	200
Sediments	AsO_3^{3-} , AsO_4^{3-}	Dionex AS1	3.0 mM NaHCO_3^- 1.4 mM Na_2CO_3	Cond.	No interference from NO_3^- , PO_4^{3-} , SO_4^{2-} ; time of analysis, 30 min	201
Sludges	Cl^- , NO_3^- , SO_4^{2-}	Dionex AS4	2.8 mM NaHCO_3^- 2.2 mM Na_2CO_3	Cond.	Time of analysis, 12 min	202
Soil	NO_3^-	Waters C ₁₈ Radial-Pak	Tetrabutylammonium hydrogen sulfate	UV (220 nm)	Time of analysis, 6 min; compared well with colorimetric method	203
Soil, plants	Total S	Dionex AS3	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	Time of analysis, 8 min; compared well with methylene blue method; R.S.D. 0.3-6.5%	204

The chief advantage of IC in plant nutrition studies is its ability for rapid analysis of many anions including intracellular organic acids. Little sample preparation is required for these analyses. Deproteinised plant extracts (obtained routinely by boiling) avoid column contamination. Other advantages of IC in the anion analysis of plant extracts have been described recently by Grunau and Swiader¹⁹⁵. However, they emphasized the need for precaution such as the use of membrane filters for the removal of interfering extractables since some of these extractables may cause column poisoning. Various digestion methods have been reported including wet and dry procedures^{194,195,197}.

4. AIRBORNE MATERIALS

IC is a valuable tool for the determination of ions of various airborne samples such as ambient air, aerosols and dust (Table 3). The sample species may be gases including NO_x , SO_x or mists such as H_2SO_4 that have been emitted from anthropogenic sources. Airborne samples also include non-volatile salts and inorganic ions sorbed on particulates such as fly ash or dust.

An airborne sample must first be dissolved in a solvent before it can be analyzed by IC. But this is not always so straightforward. For example, if SO_2 is collected, it must first be converted to an ionic form such as SO_4^{2-} (refs. 208,209) before the IC sample analysis can be performed. Peroxybenzoylnitrate is determined by hydrolysis of the sample into the benzoate anion which is then determined by IC²⁰⁶. Sulfuryl fluoride is hydrolyzed to F^- for the IC analysis²⁰⁷.

Collection of airborne samples is performed with an impinger or sampling train where the gas is sparged through a collection solvent. Frequently, the impinger solvent contains a reagent to convert the sample into the ionic form, e.g., NO_2 to NO_3^- , SO_2 to SO_4^{2-} .

Impingers containing solvents may not be convenient for field investigations or for monitoring worker exposure. Solvent-free systems are often preferred. Much of the recent work describes solid trapping samplers. Gases are passed through the sampler and then desorbed with a solvent. The sampler may be an activated carbon cartridge or flow-through denuder. Although activated charcoal may be used for direct sampling, frequently the support is coated with a chemical to make it more reactive with the target species, i.e., triethanolamine-coated supports to collect SO_2 and NO_2 (refs. 208–210).

Airborne particulates can be dry-filtered out of the air and extracted with a solvent before analysis. Frequently, water or the IC eluent is used as the extraction solvent.

Once the sample is collected, the chromatography becomes fairly straightforward. Tsitouridou *et al.*²³⁵ reported a comprehensive study on eluent and column selection for determination of more than 15 different ions in aerosol and fog water samples. Anion-exchange chromatography is most commonly used for anion samples, but ion-exclusion chromatography is becoming popular for weak inorganic and organic acid samples²¹⁷.

Much of the work on airborne samples involves the determination of Cl^- , NO_3^- and SO_4^{2-} . However, as indicated in Table 3, there are considerable studies conducted on other ions of interest. For example, determination of ammonia^{211–213}, amines²¹¹

TABLE 3
APPLICATIONS OF IC IN THE ANALYSIS OF AIRBORNE SAMPLES

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Air	Peroxybenzoylnitrate	Dionex anion exchange	2.6 mM NaHCO ₃ - 3.0 mM Na ₂ CO ₃	UV (224 nm)	Potential interference by C ₆ H ₅ CHO and C ₆ H ₅ COOH; LOD, 0.03 µg l ⁻¹	206
Air	Sulfurylfluoride	Dionex AS-1	3.5 mM Na ₂ CO ₃ - 4.0 mM NaOH	Cond.	Determination based upon hydrolysis with detection of F ⁻ and SO ₄ ²⁻ ; recovery, 95.8%; R.S.D. 11%	207
Air	Sulfur dioxide nitrogen dioxide	Dionex AS-4A	8.0 mM Na ₂ CO ₃ - 3.0 mM NaHCO ₃	Cond.	Trapped on Sep-Pak C impregnated with triethanolamine-potassium hydroxide; R.S.D. 2.4-5.3%; recovery, 85-105%	208
Air	Sulfur dioxide nitrogen dioxide	Dionex AS-1	3.0 mM NaHCO ₃ - 2.4 mM Na ₂ CO ₃	Cond.	Trapped on triethanolamine impregnated with molecular sieve; recovery, 85%	209
Air	Nitrogen dioxide	Dionex AS-4A	2.0 mM Na ₂ CO ₃ - 0.7 mM NaHCO ₃	Cond.	Trapped on Sep-Pack C ₁₈ impregnated with triethanol- amine; R.S.D. 2.7%; (84 ng ml ⁻¹), recovery, 87-94%	210
Air	Ammonia and amines	Dionex cation exchange	HCl-HNO ₃	Cond.	Trap ammonia/amines by oxalic acid-glycerol treated filter paper; LOD, 8.4 ng ml ⁻¹ NH ₃ , 7.9-12.1 ng ml ⁻¹ amines	211
Air	Ammonia	Dionex cation exchange	5.0 mM HCl	Cond.	LOD, 10 µmol l ⁻¹ ; R.S.D. <3%; compares favorably to colorimetric method	212
Auto exhaust	NH ₃ , alkylamines	Dionex cation exchange	2.5 mM HNO ₃	Cond.	Agreed well with spectroscopy at > 10 µg ml ⁻¹ NH ₃	213

Air	F ⁻	Dionex anion exchange	0.3 mM NaHCO ₃	Cond.	Al did not interfere; LOD, 0.1 µg ml ⁻¹	214
Air	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ acetate, formate, oxalate	Dionex AS-3	2.2 mM Na ₂ CO ₃ 2.8 mM NaHCO ₃	Cond.	Components of copper patina	215
Air	Formaldehyde	Dionex anion exchange	5.0 mM Na ₂ B ₄ O ₇	Cond.	Charcoal adsorption; detected as HCOO ⁻ ; recovery, 99%; R.S.D. 8%	216
Air	Di- and monomethyl sulfate	Dionex anion exchange	3.0 mM Na ₂ CO ₃ 2.6 mM NaOH	Cond.	Collected on filter pack adsorbent bed; LOD, 2-10 nmol	218
Air	N ₃ ⁻	Dionex AS2	3.0 mM NaHCO ₃ 2.4 mM Na ₂ CO ₃	Cond.	Air bag inflator effluent; R.S.D. 2.5%; linear range, 0.1-10 µg ml ⁻¹	219
Air	N ₃ ⁻	Dionex AS1	2.0 mM NaOH- 2.4 mM Na ₂ CO ₃	Cond.	Air bag inflator effluent; Br and adipate cause interference; R.S.D. 2.5%; LOD. 0.5 µg ml ⁻¹ (100 µl)	220
Air	HCHO, CH ₃ CHO	Dionex AS1	1.5 mM NaHCO ₃ 5.0 mM Na ₂ B ₄ O ₇ 6.0 mM Na ₂ CO ₃	Cond.	Conversion to formate; R.S.D. 2.6%; LOD, 0.5 ng ml ⁻¹	221
Air	AsO ₃ ³⁻ , AsO ₄ ³⁻ , monomethyl arsonate, dimethyl arsinite	Dionex anion exchange	2.4 mM NaHCO ₃ 1.9 mM Na ₂ CO ₃ 1.0 mM Na ₂ B ₄ O ₇	AAS	R.S.D. 11%; LOD < 10 ng ml ⁻¹ ; time of analysis, 10 min.	222
Air	SO ₂	Dionex anion exchange	3.0 mM Na ₂ CO ₃ 3.0 mM NaHCO ₃	Cond.	Conversion to SO ₄ ²⁻ ; recovery 83-98%; R.S.D. 2.4-7.5%; LOD, 6.0 µg ml ⁻¹	223
Air	SO ₂	Dionex AS1	NaHCO ₃ -Na ₂ CO ₃	Cond.	Impinger, H ₂ O ₂ solution with conversion to SO ₄ ²⁻	224
Air	NO ₂ , NO _x	Dionex AS4	0.20 mM Na ₂ CO ₃ 0.75 mM NaHCO ₃	Cond.	Collected in alkaline KMnO ₄ and converted into NO ₂ ⁻ ; LOD, 13 mg NO _x /m ³	225
Air	NO	Dionex AS3	3.0 mM NaHCO ₃ 2.4 mM Na ₂ CO ₃	Cond.	LOD, 0.25 µg m ⁻³ ; R.S.D. 5.7%	226

(Continued on p. 234)

TABLE 3 (*continued*)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Air	H_2SO_4 mist	Dionex anion exchange	4.0 mM Na_2CO_3^- 4.0 mM NaHCO_3	Cond.	Collected on PTFE filter, extracted with benzaldehyde; interference by benzoic acid; LOD, 0.005 $\mu\text{g m}^{-3}$	227
Air	Cl_2 , ClO_2	Dionex AS3	Na_2CO_3^- - NaHCO_3	Cond.	Time of analysis, 25 min; R.S.D. 1.4-7.3%; LOD, 0.02 $\mu\text{g ml}^{-1}$ ClO_2 and 0.06 $\mu\text{g ml}^{-1}$ Cl_2	228
Air	HCOO^- , CH_3COO^-	Dionex AS1	5.0 mM $\text{Na}_2\text{B}_4\text{O}_7$	Cond.	LOD, 0.2 $\mu\text{g m}^{-3}$ for formic acid and formaldehyde and 0.4 $\mu\text{g m}^{-3}$ for acetic acid and acetaldehyde	229
Air	ClCH_2COOH	Dionex anion exchange	1.5 mM NaHCO_3	Cond.	R.S.D. 1.7%; no interference by glyclic acid, acetic acid, dichloroacetic acid, F^- and Cl^- compounds or water vapor	230
Air	Ammonium sulfamate	Dionex anion exchange	3.0 mM Na_2CO_3^- 2.0 mM NaHCO_3	Cond.	R.S.D. 1.3-3.5%; retention time, 3.4 min; recovery 94-100%; LOD, 0.9 mg m^{-3} (90-l sample)	231
Air	SO_4^{2-} , CO_3^{2-} , NO_3^-	(i) Dionex 35311 (ii) Dionex 30890	(i) 1.5 mM Na_2CO_3^- 1.9 mM NaHCO_3 (ii) 70 mM Na_2CO_3	Cond.	Samples collected in alkaline KMnO_4 ; recovery, 99-102%; R.S.D. 0.4-4.6%	232
Air	HCOO^- , CH_3COO^- , $\text{CH}_3\text{CH}_2\text{COO}^-$, Cl^- , NO_2^- , NO_3^- , SO_2	Dionex AS2	1.0 mM Tridecafluoro- heptanoic acid, 1% isopropanol	Cond.	Recovery, 95-98%	64
Air	Chloroacetyl chloride	Dionex AS1	1.5 mM NaHCO_3	Cond.	Recovery, 99%; R.S.D. 7.2-8.8%; LOD, 0.01 $\mu\text{g ml}^{-1}$	233

Air	Cl^- , NO_3^- , SO_4^{2-}	Vydac 302 IC	1.6 mM Salicylate	UV and fluorometry	234
Aerosol	(i) Cl^- , NO_3^- , NO_3^- SO ₄ ²⁻ (ii) Na^+ , NH_4^+ , K ⁺ (iii) F ⁻ , glycolate, lactate, formate, formic acid	Wescan anion/R	(i) 4.0 mM Potassium hydrogenphthalate (ii) HNO ₃ (iii) H ₂ SO ₄	(i) Cond. (ii) Cond. (iii) Cond.	236 Portable IC, LOD, 0.02– 0.15 mg l ⁻¹
Aerosol	NH ₄ ⁺	Dionex cation exchange	6.0 mM HNO ₃	Cond.	R.S.D. 0.9–2.9%; time of analysis, 10 min; compared well with ion selectivity method
Paint aerosol	Cr(VI)	Dionex AS5	0.5 mM NaOH– 7.0 mM Na ₂ CO ₃	Cond.	LOD, 0.1 $\mu\text{g ml}^{-1}$; R.S.D. 4.3%; time of analysis, 8 min
Aerosol	SO ₄ ²⁻ , NO ₃ ⁻	Dionex AS1	3.0 mM NaHCO ₃ ⁻ 0.9 mM Na ₂ CO ₃	Cond.	Br ⁻ and PO ₄ ³⁻ interfered with analysis; time of analysis, 20 min; R.S.D. 1–3%
Aerosol	SO ₄ ²⁻	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	LOD, 0.5 $\mu\text{g m}^{-3}$
Aerosol	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Vydac 302	5.0 mM Potassium hydrogen- phthalate	Cond.	Time of analysis, 7 min
Aerosol	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	Dionex AS4	2.8 mM NaHCO ₃ ⁻ 2.2 mM Na ₂ CO ₃	Cond.	LOD, 5 ng m ⁻³ ; time of analysis, 8 min
Aerosol	F ⁻ , Cl ⁻ , (NO ₃ ⁻ , NO ₃ ⁻ , SO ₄ ²⁻)	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	Time of analysis, 20 min
Flue dust	AsO ₄ ³⁻ , SO ₄ ²⁻	Dionex anion exchange	3.5 mM Na ₂ CO ₃ ⁻ 2.6 mM NaOH	Cond.	No detection of AsO ₃ ³⁻ ; LOD, 1.8 $\mu\text{g ml}^{-1}$ AsO ₄ ²⁻ ; oxidation of SO ₃ ²⁻ to SO ₄ ²⁻ by addition of FeCl ₃ or CuCl ₂
Flue dust	AsO ₄ ³⁻ , AsO ₃ ³⁻ (Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻)	Dionex AS1	3.5 mM Na ₂ CO ₃ ⁻ 2.6 mM NaOH	Cond.	Time of analysis, 30 min
Flue gas	SO ₃ ²⁻ , SO ₄ ²⁻ , H ₂ SO ₄ miss	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	SO ₃ ²⁻ and SO ₄ ²⁻ are detected simultaneously

(Continued on p. 236)

TABLE 3 (*continued*)

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Flue gas	NO _x , SO _x	Toya Soda IC-Anion-PW	0.7 mM Gluconate-0.7 mM boron-0.1 mM KH ₂ PO ₄ -EDTA	UV-VIS-Cond.	Ions were determined from flue gas scrubbing	246
Flue gas	SO ₃ ²⁻ , SO ₄ ²⁻	Dionex anion exchange	9.0 mM NaHCO ₃ ⁻ 7.2 mM Na ₂ CO ₃	Cond.	IC method was used to determine sulfoxyl ions in flue gas desulfurization solutions	247
Flue gas	Hydrogen halides and halogens	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃	Cond.	No SO ₂ or NO _x interference; recovery 99.4%; LOD, low µg ml ⁻¹	248
Flue gas	Cl ⁻ , SO ₃ ²⁻ , SO ₄ ²⁻ , Ca ²⁺ , Mg ²⁺ , Na ⁺ , K ⁺ , NH ₄ ⁺	(i) Dionex anion exchange (ii) cation exchange	(i) 3.0 mM NaHCO ₃ ⁻ 2.4 mM Na ₂ CO ₃ (ii) 5.0 mM HNO ₃	Cond.	Time of analysis, 30 min	249
Combustion gases	F ⁻ , Cl ⁻ , NO _x , SO ₃ ²⁻ , SO ₄ ²⁻ , Br ⁻ , HPO ₄ ²⁻	Dionex anion exchange	3.0 mM NaHCO ₃ ⁻ 4.0 mM Na ₂ CO ₃	Cond.	Column temperature critical to obtain good resolution	250

Dusts, fly ash	F^- , Cl^- , NO_3^- , NO_2^- , I^- , SO_4^{2-} , PO_4^{3-}	Dionex anion exchange	3.0 mM NaHCO ₃ 2.4 mM Na ₂ CO ₃	Cond.	Time of analysis, 16 min; R.S.D. 0.12–9.99%; compared well with ion selective electrodes, potentiometry, AAS, ICP	251
Coal fly ash	F^- , Cl^- , NO_3^- , SO_4^{2-}	Dionex anion exchange	3.0 mM NaHCO ₃ 2.4 mM Na ₂ CO ₃	Cond.	R.S.D. < 3%; LOD, 0.05–150 $\mu\text{g ml}^{-1}$; unable to detect PO_4^{3-}	252
Diesel exhaust	Formic acid	Dionex ICE	5.0 mM Na ₂ B ₄ O ₇	Cond.	Time of analysis, 11 min; peak height linear from 0.1–4 $\mu\text{g ml}^{-1}$; LOD, 0.05 $\mu\text{g ml}^{-1}$ (100 μl sample)	217
Fuel combustion products	F^- , Cl^- , PO_4^{3-} , NO_2^- , NO_3^- , SO_4^{2-} , Mg^{2+} , Ca^{2+} , Na^+ , NH_4^+ , K^+	Dionex AS1	3.0 mM NaHCO ₃ 2.4 mM Na ₂ CO ₃	Cond.	Time of analysis, 20 min; compares to ASTM methods with greater sensitivity	253
Atmospheric pollutants	NO_2^- , NO_3^- , SO_4^{2-}	IC-PAK anion	Potassium hydrogenphthalate vs. borate–gluconate	Cond.-UV (284 nm)	IC compared well to colorimetric method for NO_2^- ; time of analysis, 16 min; LOD by UV for SO_4^{2-} , 5 ng (50 μl sample)	254

TABLE 4
APPLICATIONS OF IC IN THE ANALYSIS OF FOSSIL FUELS

Sample	Ion	Analytical column	Eluent	Detector	Comments	Ref.
Fuels, coal	F^- , Cl^- , NO_3^- , PO_4^{3-} , Br^- , NO_3^- , SO_4^{2-}	Dionex anion exchange	Na_2CO_3 - NaHCO_3	Cond.	Compared well to ASTM methods; LOD, $2-10 \mu\text{g ml}^{-1}$	255
Fuels	Total S (F^- , Cl^- , NO_3^- , SO_4^{2-})	Dionex anion exchange	2.4 mM NaCO_3^- 3.0 mM NaHCO_3	Cond.	Compared well to ASTM methods; R.S.D. 0.7%; LOD, $2 \mu\text{g ml}^{-1}$ SO_4^{2-}	256
Fuel oil	Total S	Dionex AS1	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	Compared well with titrimetry	257
Fuel oil	Total S	Dionex AS1	3.0 mM NaHCO_3^- 2.4 mM Na_2CO_3	Cond.	Compared well with X-ray fluorescence and titrimetry; LOD, 0.007%; R.S.D. 2.7%; time of analysis, 25 min	258
Crude oil	Total S (Cl^- , NO_3^- , SO_4^{2-})	Waters IC-Pak	Borate-gluconate-acetonitrile-glycerol	Cond.	Recovery, 95.7-102.2%; R.S.D., 1.0-2.9%; LOD, 0.5 $\mu\text{g kg}^{-1}$; time of analysis, 15 min	259
Coal, ash, geological materials	F^-	Dionex AS3	1.5 mM NaHCO_3	Cond.	LOD, 1 ng; R.S.D. 4.9%; compared well with ion selective electrode	260
Coal	Total Cl ⁻	Wescan 260-001	Potassium hydrogenphthalate	Cond.	Extraction with dimethyl sulphoxide; converted to Cl ⁻	261
Coal	Cl^- , NO_3^- , NO_2^- , SO_4^{2-}	SAX 1	4.0 mM Na_2CO_3^- 4.0 mM NaHCO_3	Cond.	Precision not satisfactory for NO_2^- , NO_3^- ; time of analysis, 10 min	262
Coal	Ag, Al, Ba, Cd, Cu, Fe, Mg, Mn, Ni, Pb, Si, Sn, Ti, V, Zn	100 Å μ Styragel	Pyridine (chloroform, tetrahydrofuran)	ICP-AES	LOD, 3-530 ng ml^{-1} ; size-exclusion chromatography; multiple peaks of Fe	263

and alkylamines²¹³ have been reported. Acetate, formate and oxalate²¹⁵, sulfamate²³¹, monochloroacetic acid²³⁰, chloroacetyl chloride²³³, formic acid^{64,217,235}, dimethyl- and methyl-sulfate²¹⁸, azide^{219,220}, formaldehyde²¹⁶ and acetaldehyde²²¹ have also been monitored.

5. FOSSIL FUELS

The elemental composition of fossil fuels such as coal and various petroleum products is an important environmental consideration in the use of these energy sources. IC can be used to determine the fuel elemental composition for halogens, N, S, P and various metals. The samples may first be combusted in an oxygen bomb or a furnace, the gases are absorbed in water and the ions are determined by IC. In other cases, the sample is simply extracted with a solvent. Dimethyl sulfoxide has been used to leach Cl⁻ from a number of coal samples²⁶¹. Water extract of coal has been used by Honma *et al.*²⁶² for the determination of Cl⁻, NO₂⁻, NO₃⁻ and SO₄²⁻. Recently Burns *et al.*²⁶⁴ found IC to be very attractive for the separation of a number of radionuclides of elements such as plutonium, americium, curium, uranium and thorium in radiochemical wastes. Table 4 lists the applications of IC in the analysis of fossil fuels.

6. CONCLUSIONS

New analytical developments in IC have allowed the simultaneous determination of ions from complex matrices at trace levels. IC can be tailored to be selective for specific target solutes in the presence of other matrix ions, *i.e.*, SeO₄²⁻ vs. SO₄²⁻ (ref. 173), SeO₃²⁻ vs. Cl⁻ (ref. 174) and AsO₃³⁻ vs. PO₄³⁻ (ref. 177). Much of the IC applications involve suppressed systems marketed by Dionex (Sunnyvale, CA) but recent advances have now become available with non-suppressed systems (single column ion chromatography, SCIC).

Important parameters in optimizing the separation and detection of target solutes include choice of eluent, concentration of the mobile phase, working pH, column selection and choice of detectors. Among the most popular eluents, low ionic strength aromatic organic acids (phthalate, benzoate and citrates) and mineral acids have been used as the mobile phase in SCIC. Stationary phases include macroporous styrenedivinylbenzene resins, quaternized silica and polymethacrylate gels. Conductivity is frequently used as the mode of detection in IC but other detectors include absorbance (UV or visible light), fluorescence, AAS, coulometry and amperometry. Electrochemical detection is popular because of its high selectivity and its ability to detect ultra-trace levels.

New developments in IC do not simply involve "pick and choose" of each component (mobile phase, column and detector), but require extensive knowledge in determining the compatibility of these components in developing a useful analytical system. The pK_a of the target ion of interest must be known as well as the presence of potential interfering substances. Column selection is based on the upper and lower pH limit of the mobile phase to optimize separation.

Among the advantages of IC are: (i) its relatively high sensitivity with detection limits in the low $\mu\text{g ml}^{-1}$ (conductivity, UV/VIS, AAS), low ng ml^{-1} (amperometry)

and pg ml^{-1} range (fluorometric detection); (ii) simultaneous determination of several species and other elements, e.g., separation and determination of SO_3^{2-} , SO_4^{2-} and $\text{S}_2\text{O}_3^{2-}$ (ref. 108); (iii) IC is non-destructive and fractions can be collected for further confirmation; (iv) the cost of IC is relatively low (the system can be modified easily by combining different components to accommodate changes in analytical requirements); (v) sample preparation is often minimal. IC can be used in routine analysis of environmental samples and operated after minimal instruction.

7. SUMMARY

Ion chromatography (IC) is going through rapid growth and popularity because of its usefulness in the applied environmental field. This review includes recent IC applications in rainwater, groundwater, surface water, wastewater, drinking water, fog samples, ice, snow, soil, sediments, sludge, plants, air, exhaust, aerosols, flue dust, fly ash, fuel oil and coal. A major emphasis of this review is on speciation of ions.

REFERENCES

- 1 H. Small, T. S. Stevens and W. C. Bauman, *Anal. Chem.*, 47 (1975) 1801.
- 2 E. Sawicki, J. D. Mulik and E. Wittgenstein (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 1, Ann Arbor Sci. Publ., Ann Arbor, MI, 1978.
- 3 J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979.
- 4 D. T. Gjerde and J. S. Fritz, *Ion Chromatography*, Hüthig, Mamaroneck, NY, 2nd ed., 1987.
- 5 R. Smith, *Ion Chromatography Applications*, CRC Press, Boca Raton, FL, 1988.
- 6 J. Weiss, *Handbook of Ion Chromatography*, Dionex, Sunnyvale, CA, 1986.
- 7 J. G. Tarter, *Ion Chromatography*, Marcel Dekker, New York, 1987.
- 8 F. C. Smith and R. C. Chang, *The Practice of Ion Chromatography*, John Wiley, New York, 1983.
- 9 H. Small, *React. Polym. Ion Exch. Sorbents*, 7 (1988) 73.
- 10 H. W. Stuurman, *Int. Lab.*, 18, No. 3 (1988) 34.
- 11 F. Marcenac, *Analisis*, 16 (1988) 57.
- 12 J. Weiss, *Fresenius' Z. Anal. Chem.*, 327 (1987) 451.
- 13 J. Behnert, P. Behrend and A. Kipplinger, *Inorganic Analytical Chemistry*, Vogel-Verlag, Weierzburg, 1987, p. 32.
- 14 T. Nomura, *Zairyo Gijutsu*, 5 (1987) 472.
- 15 H. Small, in J. R. Lawrence (Editor), *Trace Analysis*, Vol. 1, Academic Press, New York, NY, 1981, p. 267.
- 16 M. D. Gaul, G. W. Boyes, G. Kookootsedes and A. M. Waldern, *Sampe J.*, Nov./Dec. (1987) 22.
- 17 J. Weiss, *Fresenius' Z. Anal. Chem.*, 327 (1987) 25.
- 18 G. Schmuckler, *J. Liq. Chromatogr.*, 10 (1987) 1887.
- 19 R. B. Rubin and S. S. Heberling, *Int. Lab.*, (1987) 54.
- 20 J. S. Fritz, *Anal. Chem.*, 59 (1987) 335A.
- 21 R. Hatano, *Nippon Dojo Hiryogaku Zasshi*, 57 (1986) 421.
- 22 R. Matas Docampo, *Tec. Lab.*, 10 (1986) 528.
- 23 I. S. Krull, *Life Sci. Res. Rep.*, 33 (1986) 579.
- 24 Z. Shang, J. Wang and L. Zhou, *Huaxue Shiji*, 8 (1986) 339.
- 25 R. A. Cochrane, *Spec. Publ.-R. Soc. Chem.*, 61 (1986) 197.
- 26 C. Jansen and J. Weiss, *Umwelt.*, 5 (1986) 411.
- 27 T. H. Jupille and D. T. Gjerde, *J. Chromatogr. Sci.*, 24 (1986) 427.
- 28 G. O. Franklin, *Am. Lab. (Fairfield, Conn.)*, 17 (1985) 65.
- 29 P. R. Haddad and A. L. Heckenberg, *J. Chromatogr.*, 300 (1984) 357.
- 30 P. McFadyen, *International Labmate*, 9 (1984) 21.

- 31 H. Small, *Anal. Chem.*, 55 (1983) 235A.
- 32 O. A. Shpigun and Yu. A. Zolotov, *Zavod. Lab.*, 48 (1982) 4.
- 33 C. Pohlhardt, *S. Afr. Tydskr. Chem.*, 33 (1980) 87.
- 34 J. C. MacDonald, *Am. Lab. (Fairfield, Conn.)*, 11 (1979) 45.
- 35 R. Bogoczek and G. Miemus, *Przem. Chem.*, 59 (1980) 471.
- 36 F. C. Smith, R. C. Chang, *CRC Crit. Rev. Anal. Chem.*, 9 (1980) 197.
- 37 K. Oikawa, *Bunseki*, 8 (1980) 531.
- 38 A. Jardy and R. Rosset, *Analisis*, 7 (1979) 259.
- 39 Y. Tsuchitani, *Bunseki*, 9 (1979) 603.
- 40 T. Nomura, *Kasaku to Kosyo, Sci. Ind. (Osaka)*, 52 (1978) 448.
- 41 R. A. Nadkarni and J. M. Brewer, *Am. Lab. (Fairfield, Conn.)*, 19 (1987) 50.
- 42 E. J. Roekens, P. M. Otten and R. E. van Grieken, *Chem. Mag.*, 13 (1987) 719.
- 43 *Application Note, 305: Ion Analyzer Bulletin 110*, Wescan, Deerfield, IL, 1987.
- 44 W. H. McDowell and S. Stedman, *First Ion Chromatography Forum, Boston, MA, Sept. 25, 1988*, Abstract No. 21.
- 45 J. Veisserik, K. Punning and E. Nilson, *Symp. Biol. Hung.*, 31 (1986) 57.
- 46 V. Pihl, Ya. O. Penchuk, L. Margna and K. Ilmoja, *Uch. Zap. Tartu. Gos. Univ.*, 743 (1986) 176.
- 47 V. K. Kondawar, V. R. Bhave, N. Thakkar and P. L. Muthal, *Indian J. Environ. Health*, 28 (1986) 324.
- 48 S. Mou and W. Ye, *Huanjing Huaxue*, 5 (1986) 68.
- 49 B. A. Colenutt and P. J. Trenchard, *Environ. Pollut. Ser. B*, 10 (1985) 77.
- 50 A. J. Lipski and C. J. Vairo, *Can. Res.*, 13 (1980) 45.
- 51 W. E. Rich and R. A. Wetzel, *Actual. Chim.*, 6 (1980) 51.
- 52 R. Wetzel, *Environ. Sci. Technol.*, 13 (1979) 1214.
- 53 W. E. Rich and R. A. Wetzel, in D. Schuetze (Editor), *Monitoring of Toxic Substances (ACS Symposium Series, No. 94)*, American Chemical Society, Washington, DC, 1979, p. 233.
- 54 K. Oikawa, *PPM*, 9 (1978) 52.
- 55 R. K. Skogerboe, *Toxicol. Environ. Chem. Rev.*, 2 (1978) 209.
- 56 W. E. Rich, *Instrum. Technol.*, 24 (1977) 47.
- 57 K. J. Bombaugh, *Water Anal.*, 3 (1984) 317.
- 58 J. L. Peschet and C. Tinet, *Eau. Ind., Nuisances*, 114 (1987) 47.
- 59 M. Lindgren, *Vatten*, 36 (1980) 249.
- 60 G. S. Pyen and D. E. Erdmann, *Anal. Chim. Acta*, 149 (1983) 355.
- 61 I. T. Urasa and S. H. Nam, *J. Chromatogr. Sci.*, 27 (1989) 30.
- 62 S. Dogan and W. Haerdi, *Chimia*, 35 (1981) 339.
- 63 S. Motomizu, I. Sawatani, T. Hironaka, M. Oshima and K. Toci, *Bunseki Kagaku*, 36 (1987) 77.
- 64 D. Brocco and R. Tappa, *J. Chromatogr.*, 367 (1986) 240.
- 65 T. Okutani and Y. Yugeta, *Bunseki Kagaku*, 34 (1985) 777.
- 66 J. A. Hern, G. K. Rutherford and G. W. vanLoon, *Talanta*, 30 (1983) 677.
- 67 J. P. Ivey and P. R. Haddad, *J. Chromatogr.*, 391 (1987) 309.
- 68 D. M. Davies and J. P. Ivey, *Anal. Chim. Acta*, 194 (1987) 275.
- 69 J. Crowther and J. McBride, *Analyst (London)*, 106 (1981) 702.
- 70 N. Hirayama and T. Kuwamoto, *J. Chromatogr.*, 457 (1988) 415.
- 71 D. C. Bogen and S. J. Nagourney, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 319.
- 72 S. Y. Tyree, Jr., J. M. Stouffer and M. Bollinger, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 295.
- 73 S. Kadowaki, *Kogai to Taisaku*, 23 (1987) 1167.
- 74 T. Hironaka, M. Oshima and S. Motomizu, *Bunseki Kagaku*, 36 (1987) 503.
- 75 V. Cheam and A. S. Y. Chau, *Analyst (London)*, 112 (1987) 993.
- 76 R. Hill and K. H. Leiser, *Fresenius' Z. Anal. Chem.*, 327 (1987) 165.
- 77 D. C. Gan and J. G. Tarter, *J. Chromatogr.*, 404 (1987) 285.
- 78 D. C. Schroeder, *J. Chromatogr. Sci.*, 25 (1987) 405.
- 79 F. G. P. Mullins, *Analyst (London)*, 112 (1987) 665.
- 80 G. Saito, T. Kimura, M. Kojima and F. Hayase, *Nippon Dojo Hiryogaku Zasshi*, 58 (1987) 1.
- 81 T. Okutani and M. Tanaka, *Bunseki Kagaku*, 36 (1987) 169.
- 81 Y. Hoshika, N. Murayama and G. Muto, *Bunseki Kagaku*, 36 (1987) 174.

- 83 T. Iwachido, K. Ishimaru and S. Motomizu, *Anal. Sci.*, 4 (1988) 81.
- 84 F. R. Nordmeyer, L. D. Hansen, D. J. Eatough, D. K. Rollins and J. D. Lamb, *Anal. Chem.*, 52 (1980) 852.
- 85 D. L. Smith and J. S. Fritz, *Anal. Chim. Acta*, 204 (1988) 87.
- 86 J. D. Lamb, L. D. Hansen, G. G. Patch and F. R. Nordmeyer, *Anal. Chem.*, 53 (1981) 749.
- 87 J. R. Dean, *Analyst (London)*, 114 (1989) 165.
- 88 P. Jones, L. Ebdon and T. Williams, *Analyst (London)*, 113 (1988) 641.
- 89 K. Tanaka, T. Ishizuka and H. Sunahara, *J. Chromatogr.*, 177 (1979) 21.
- 90 V. T. Wee and J. M. Kennedy, *Anal. Chem.*, 54 (1982) 1631.
- 91 D. Chakraborti and K. J. Irgolic, in T. D. Lekkas (Editor), *Heavy Metal Environment, 5th International Conference, Athens, 1985*, Vol. 2, CEP Consultants, Edinburgh, 1985.
- 92 H. Itoh and Y. Shinburi, *Bunseki Kagaku*, 29 (1980) 239.
- 93 J. P. Romano, *J. Anal. Purif.*, 2 (1987) 68.
- 94 K. Ito and H. Sunahara, *Bunseki Kagaku*, 37 (1988) 292.
- 95 P. R. Haddad and R. C. Foley, *Anal. Chem.*, 61 (1989) 1435.
- 96 C. Erkelens, H. A. H. Billiet, L. De Galan and E. W. B. De Leer, *J. Chromatogr.*, 404 (1987) 67.
- 97 G. Schwedt, B. C. Seo, S. Dreyer and E. U. Ruhdel, *LaborPraxis*, 10 (1986) 1308.
- 98 J. A. Oppenheimer, A. D. Eaton and P. H. Kreft, *EPA/600/2-84/190*, U.S. Environmental Protection Agency, Cincinnati, 1984.
- 99 G. S. Pyen and M. J. Fishman in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 235.
- 100 *Environmental Application Note, WB0927*, Waters, Milford, MA, 1987.
- 101 M. Legrand, M. De Angelis and R. J. Delmas, *Anal. Chim. Acta*, 156 (1984) 181.
- 102 C. Saigne, S. Kirchner and M. Legrand, *Anal. Chim. Acta*, 203 (1987) 11.
- 103 Y. A. Zolotov, O. A. Shpigun, Yu. E. Pazukhina and I. N. Voloshik, *Int. J. Environ. Anal. Chem.*, 31 (1987) 99.
- 104 N. S. Simon, *Anal. Lett.*, 21 (1988) 319.
- 105 J. A. Mosko, *Anal. Chem.*, 56 (1984) 629.
- 106 M. E. Potts and T. A. Potas, *J. Chromatogr. Sci.*, 23 (1985) 411.
- 107 M. Nonomura, *Anal. Chem.*, 59 (1987) 2073.
- 108 R. E. Poulsom and H. M. Borg, *J. Chromatogr. Sci.*, 25 (1987) 409.
- 109 L. W. Green and J. R. Woods, *Anal. Chem.*, 53 (1981) 2187.
- 110 D. L. Wilson, H. B. Durham and R. C. Thurnau, *LC GC*, 4 (1986) 578.
- 111 S. Y. Tyree Jr. and M. A. Bynum, *Limnol. Oceanogr.*, 29 (1984) 1337.
- 112 L. R. Goodwin, D. Francom, A. Urso and F. P. Dieken, *Anal. Chem.*, 60 (1988) 216.
- 113 K. Punning, T. Somer and J. Veisserik, *Symp. Biol. Hung.*, 34 (1986) 541.
- 114 M. Nonomura, *Met. Finish.*, December (1987) 15.
- 115 H. Mehra and W. T. Frankenberger, Jr., *Microchem. J.*, 41 (1990) in press.
- 116 R. P. Lash and C. J. Hill, *J. Liq. Chromatogr.*, 2 (1979) 417.
- 117 F. J. Trujillo, M. M. Miller, R. K. Skogerboe, H. E. Taylor and C. L. Grant, *Anal. Chem.*, 53 (1981) 1944.
- 118 C. J. Hill and R. P. Lash, *Anal. Chem.*, 52 (1980) 24.
- 119 A. Yu. Zolotov, E. Yu. Pazukhina, O. A. Shpigun, I. N. Voloshchik and M. L. Litvia, *Dokl. Akad. Nauk./SSSR*, 297 (1987) 105.
- 120 D. Yang and Hu, *Turang (Nanjing)*, 18 (1986) 273.
- 121 W. Merz and J. Oldeweme, *Vom Wasser*, 69 (1987) 95.
- 122 D. Cao and Y. Luo, *Sepu*, 5 (1987) 189.
- 123 J. Krol, A. L. Heckenberg, W. Jones and P. Jandik, *Presented at 39th Pittsburgh Conference on Analytical Chemistry and Applied Spectroscopy, New Orleans, February 22-25, 1988*.
- 124 *Technical Note TN 24*, May 24, Dionex, Sunnyvale, CA, 1987.
- 125 L. Xiang, Z. Luo, S. Wang and C. Yang, *Shanghai Huanjing Kexue*, 6 (1987) 34.
- 126 T. Darimont, G. Schulze and M. Sonneborn, *Fresenius' Z. Anal. Chem.*, 314 (1983) 383.
- 127 N. Chavret and J. Hubert, *J. Chromatogr.*, 469 (1989) 329.
- 128 T. Okada, *Bunseki Kagaku*, 36 (1987) 702.
- 129 H. J. Kim and Y. K. Kim, *Anal. Chem.*, 61 (1989) 1485.
- 130 W. Wildman, D. Scarchilli and L. Jagoe, *J. Anal. Purif.*, 3 (1988) 50.
- 131 B. W. Smee, G. E. M. Hall and D. J. Koop, *J. Geochem. Explor.*, 10 (1978) 245.

- 132 M. Cooke, *J. High Resolut. Chromatogr. Chromatogr. Commun.*, 6 (1983) 383.
 133 G. Marko-Varga, I. Csiky and J. A. Jonsson, *Anal. Chem.*, 56 (1984) 2066.
 134 H. Sato, *Anal. Chim. Acta*, 206 (1988) 281.
 135 Yu. A. Zolotov and O. A. Shpigun, L. A. Bubchikova and E. A. Sedel'nikova, *Dokl. Akad. Nauk SSSR*, 263 (1982) 889.
 136 Y. Shibata, M. Morita and K. Fuwa, *Analyst (London)*, 110 (1985) 126.
 137 D. Togami and D. J. Hometchko, presented at *Rocky Mountain Conference, Denver, CO August 1988*.
 138 M. Ohata, A. Yamamoto, H. Araki and H. Yonezawa, *Iyakuhin Kenkyu*, 18 (1987) 760.
 139 G. Schwedt and B. Roessner, *Fresenius' Z. Anal. Chem.*, 327 (1987) 499.
 140 W. H. Ficklin, *Anal. Lett.*, 15 (1982) 865.
 141 P. Jones, L. Ebdon and T. Williams, *Analyst (London)*, 113 (1988) 641.
 142 J. S. Fritz, D. T. Gjerde and R. M. Becker, *Anal. Chem.*, 52 (1980) 1519.
 143 D. Yan and G. Schwedt, *Fresenius' Z. Anal. Chem.*, 320 (1985) 121.
 144 W. R. Jones, A. L. Heckenberg and P. Jandik, *J. Anal. Purif.*, 1 (1986) 68.
 145 D. T. Gjerde, *Report IS-T-916*, available from NTIS, *Energy Res. Abstr.*, 6 (1981) Abstr. No. 15851.
 146 S. H. Kok, K. A. Buckle and M. Wootton, *J. Chromatogr.*, 260 (1983) 189.
 147 P. J. Galvin and J. A. Cline, *Atmos. Environ.*, 12 (1978) 1163.
 148 U. Baltensperger and S. Kern, *J. Chromatogr.*, 439 (1988) 121.
 149 J. P. Ivey and D. M. Davies, *Anal. Chim. Acta*, 194 (1987) 281.
 150 D. Yan and G. Schwedt, *Fresenius' Z. Anal. Chem.*, 327 (1987) 503.
 151 H. L. Tucker and S. J. van Hook, III, presented at *Rocky Mountain Conference, Denver, CO, August 1988*.
 152 V. Cheam and E. Li, *J. Chromatogr.*, 450 (1988) 361.
 153 M. Matsumoto, *J. Jpn. Soc. Air Pollut.*, 23 (1988) 64.
 154 S. Tanaka, K. Yasue, N. Katsura, Y. Tanno, Y. Hashimoto, *Bunseki Kagaku*, 37 (1988) 665.
 155 J. Neubauer and K. G. Heumann, *Fresenius' Z. Anal. Chem.*, 331 (1988) 170.
 156 T. Nomura, Y. Hikchi and G. Nakagawa, *Bull. Chem. Soc. Jpn.*, 61 (1988) 2993.
 157 G. A. Ubom and Y. Tsuchiya, *Water Res.*, 221 (1988) 1455.
 158 E. C. V. Butler, *J. Chromatogr.*, 450 (1988) 353.
 159 T. Iwachido, K. Ishimaru, S. Motomizu, *Anal. Sci.*, 4 (1988) 81.
 160 M. Ohata, A. Yamamoto, H. Araki, H. Yonezawa, *Iyakuhin Kenkyu*, 18 (1987) 760.
 161 S. Wakida, T. Tanaka, A. Kawahara, K. Hiro, *Anal. Sci.*, 1 (1985) 355.
 162 J. L. Veuthey, J. P. Senn and W. Haerdi, *J. Chromatogr.*, 445 (1988) 183.
 163 T. H. Nguyen and L. M. Baker, *Proc. Water Qual. Technol. Conf.*, 15 (1987) 577.
 164 E. A. Stallings, L. M. Candelaria and E. S. Gladney, *Anal. Chem.*, 60 (1988) 1246.
 165 D. G. Maynard, Y. P. Kalra and F. G. Radford, *Soil Sci. Soc. Am. J.*, 51 (1987) 801.
 166 B. M. Stewart, *J. Soil Sci.*, 38 (1987) 415.
 167 U. Karlson and W. T. Frankenberger, Jr., *Soil Sci. Soc. Am. J.*, 51 (1987) 72.
 168 K. F. Nieto and W. T. Frankenberger, Jr., *Soil Sci. Soc. Am. J.*, 49 (1985) 592.
 169 P. Barak and Y. Chen, *Soil Sci. Soc. Am. J.*, 51 (1987) 257.
 170 W. A. Dick and M. A. Tabatabai, *Soil Sci. Soc. Am. J.*, 43 (1979) 899.
 171 E. G. Bradfield and D. T. Cooke, *Analyst (London)*, 110 (1985) 1409.
 172 H. Yamada, T. Hattori, S. Matuda and Y. Kang, *Bunseki Kagaku*, 36 (1987) 542.
 173 U. Karlson and W. T. Frankenberger, Jr., *J. Chromatogr.*, 368 (1986) 153.
 174 U. Karlson and W. T. Frankenberger, Jr., *Anal. Chem.*, 58 (1986) 2704.
 175 H. C. Mehra and W. T. Frankenberger, Jr., *Chromatographia*, 25 (1988) 585.
 176 S. Maketon and J. G. Tarter, *LC Mag. Liq. Chromatogr. HPLC*, 2 (1984) 124.
 177 H. C. Mehra and W. T. Frankenberger, Jr., *Soil Sci. Soc. Am. J.*, 52 (1988) 1603.
 178 R. S. Bowman, *J. Chromatogr.*, 285 (1984) 467.
 179 P. Jandik, *J. Anal. Purif.*, 3 (1988) 80.
 180 H. C. Mehra and W. T. Frankenberger, Jr., *Talanta*, 36 (1989) 889.
 181 M. Ahmad and A. Khan, *Nucleus (Karachi)*, 19 (1982) 35.
 182 H. C. Mehra and W. T. Frankenberger, Jr., *Analyst (London)*, 114 (1989) 707.
 183 H. C. Mehra and W. T. Frankenberger, Jr., *Anal. Chim. Acta*, 217 (1989) 383.
 184 M. Ahmad and A. Khan, *Nucleus (Karachi)*, 18 (1981) 29.
 185 K. F. Nieto and W. T. Frankenberger, Jr., *Soil Sci. Soc. Am. J.*, 49 (1985) 587.
 186 N. T. Basta and M. A. Tabatabai, *Soil Sci. Soc. Am. J.*, 49 (1984) 84.

- 187 N. T. Basta and M. A. Tabatabai, *American Society of Agronomy 1988 Annual Meetings, Anaheim, CA, Agronomy Abstracts*, p. 194.
- 188 P. M. Bertsch and M. A. Anderson, *Soil Sci. Soc. Am. J.*, 52 (1988) 540.
- 189 P. C. Bossle, D. J. Reutter and E. W. Sarver, *J. Chromatogr.*, 407 (1987) 399.
- 190 H. C. Mehra, K. D. Huysmans and W. T. Frankenberger, Jr., *J. Chromatogr.*, 508 (1990) in press.
- 191 H. Yamada and T. Hattori, *J. Chromatogr.*, 411 (1987) 401.
- 192 T. Ohyama, *Nippon Dojo Hiryogaku Zasshi*, 57 (1986) 503.
- 193 A. Jardy, M. Caude, A. Diop, C. Curval and R. Rosset, *J. Chromatogr.*, 439 (1988) 137.
- 194 J. Gorham, in P. A. Williams and M. J. Hudson (Editors), *Recent Developments in Ion Exchange*, Elsevier, London, 1987, p. 14.
- 195 J. A. Grunau and J. M. Swiader, *Commun. Soil Sci. Plant Anal.*, 17 (1986) 321.
- 196 C. H. Risner, *Tob. Sci.*, 30 (1986) 35.
- 197 N. T. Basta and M. A. Tabatabai, *Soil Sci. Soc. Am. J.*, 40 (1985) 76.
- 198 J. R. Wohler, F. J. Drone, J. W. Seay and J. L. Downing, *J. Anal. Purif.*, 3 (1988) 18.
- 199 R. D. Wilkes and H. H. Kock, *Fresenius' Z. Anal. Chem.*, 320 (1985) 477.
- 200 X. H. Yong and H. Zhang, *J. Chromatogr.*, 436 (1988) 107.
- 201 T. Takomatsu, M. Kawashima and M. Koyama, *Bunseki Kagaku*, 28 (1979) 596.
- 202 M. Bondonini, S. DeFulvio, F. Giorgi, P. Morsia, L. Olori and E. Veschetti, *Boll. Chim. Ig., Parte Sci.*, 35 (1984) 293.
- 203 C. C. duPreez and D. J. Laubscher, *Commun. Soil Sci. Plant Anal.*, 20 (1989) 113.
- 204 M. A. Tabatabai, N. T. Basta and H. J. Pinela, *Commun. Soil Sci. Plant Anal.*, 19 (1988) 1701.
- 205 M. Meaney, J. Mooney, M. Connor and M. R. Smith, *Anal. Proc. (London)*, 25 (1988) 63.
- 206 K. Fung and D. Grosjean, *Sci. Total Environ.*, 46 (1985) 29.
- 207 S. A. Bouyoucos, R. G. Melcher and J. R. Vaccaro, *Am. Ind. Hyg. Assoc. J.*, 44 (1983) 57.
- 208 Y. Nishihawa and K. Taguchi, *J. Chromatogr.*, 396 (1987) 251.
- 209 D. V. Vinjamoori and C. Ling, *Anal. Chem.*, 53 (1981) 1689.
- 210 Y. Nishikawa, K. Taguchi, Y. Tsujino and K. Kuwata, *J. Chromatogr.*, 370 (1986) 121.
- 211 I. Kifune and K. Oikawa, *Niigata Rikagaku*, 5 (1979) 9.
- 212 J. Rudling, B. O. Hallberg, M. Hultengren and A. Hultman, *Scand. J. Work, Environ. Health*, 10 (1984) 197.
- 213 R. B. Zweidinger, S. B. Tejada, J. E. Sigsby and R. L. Bradow, in E. Sawicki, J. D. Mulik and E. Wittgenstein (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 1, Ann Arbor Sci. Publ., Ann Arbor, MI, 1978, p. 125.
- 214 M. Oehme and H. Stray, *Fresenius' Z. Anal. Chem.*, 306 (1981) 356.
- 215 A. J. Muller and C. McCrory-Joy, *Corros. Sci.*, 27 (1987) 695.
- 216 *Application Note*, 24, Dionex, Sunnyvale, CA, 1979.
- 217 I. Bodek and K. T. Menzies, in G. Choudhary (Editor), *Chemical Hazards in the Workplace: Measurement and Control (ACS Symposium Series, No. 149)*, American Chemical Society, Washington, DC, 1981, p. 599.
- 218 L. D. Hansen, V. F. White and D. J. Eatough, *Environ. Sci. Technol.*, 20 (1986) 872.
- 219 L. C. Westwood and E. L. Stokes, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 141.
- 220 *Application Note*, 14, Dionex, Sunnyvale, CA, 1978.
- 221 J. M. Lorrain, C. R. Fortune and B. Dellinger, *Anal. Chem.*, 53 (1981) 1302.
- 222 G. R. Ricci, L. S. Shepard, G. Colovos and N. E. Hester, *Anal. Chem.*, 53 (1981) 610.
- 223 D. L. Smith, W. S. Kim and R. E. Kupel, *Am. Ind. Hyg. Assoc. J.*, 41 (1980) 485.
- 224 J. D. Mulik, G. Todd, E. Estes, R. Puckett, E. Sawicki and D. Williams, in E. Sawicki, J. D. Mulik and E. Wittgenstein (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 1, Ann Arbor Sci. Publ., Ann Arbor, MI, 1978, p. 23.
- 225 *Fed. Reg.*, 49 (189), Sept. 27, 1984, 38232-8.
- 226 P. Buttini, V. Dipalo and M. Possanzini, *Sci. Total Environ.*, 61 (1987) 59.
- 227 S. Tanaka, K. Yamanaka, K. Yamagata, Y. Komazaki and Y. Hashimoto, *Bunseki Kagaku*, 36 (1987) 159.
- 228 E. Björkhalm, A. Hultman and J. Rudling, *J. Chromatogr.*, 457 (1988) 409.
- 229 S. Tanaka, M. Iguchi, K. Yamanaka, T. Yamada, N. Nakao and Y. Hashimoto, *Bunseki Kagaku*, 36 (1987) 12.
- 230 D. W. Mason, H. K. Dillon and R. A. Glaser, *Am. Ind. Hyg. Assoc. J.*, 47 (1986) 14.

- 231 I. Bodek and R. H. Smith, *Am. Ind. Hyg. Assoc. J.*, 41 (1980) 603.
232 J. M. Margeson, J. E. Knoll and M. R. Midgett, *J. Air Pollut. Control Assoc.*, 38 (1988) 388.
233 P. R. McCullough and J. W. Worley, *Anal. Chem.*, 51 (1979) 1120.
234 S. Rapsomanikis and R. M. Harrison, *Anal. Chim. Acta*, 199 (1987) 41.
235 R. Tsitouridou and H. Puxbaum, *Int. J. Environ. Anal. Chem.*, 31 (1987) 11.
236 J. D. Mulik, E. Estes and E. Sawicki, in E. Sawicki, J. D. Mulik and E. Wittgenstein (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 1, Ann Arbor Sci. Publ., Ann Arbor, MI, 1978, p. 41.
237 D. Molina and M. T. Abell, *Am. Ind. Hyg. Assoc. J.*, 48 (1987) 830.
238 J. Mulik, R. Puckett, D. Williams and E. Sawicki, *Anal. Lett.*, 9 (1976) 653.
239 D. W. Mason and H. C. Miller, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 193.
240 W. Landolt and H. R. Moser, *Staub Reinhalt. Luft*, 45 (1985) 338.
241 G. R. Fuchs, E. Lissom, B. Schwarz and K. Bachmann, *Fresenius' Z. Anal. Chem.*, 320 (1985) 498.
242 H. Hara, K. Nagara, K. Honda and A. Goto, *Taiki Osen Gakkaishi*, 15 (1980) 380.
243 L. D. Hansen, B. E. Richter, D. K. Rollins, J. D. Lamb and D. J. Eatough, *Anal. Chem.*, 51 (1979) 633.
244 *Application Note*, 22, Dionex, Sunnyvale, CA, 1979.
245 R. Steiber and R. M. Statnick, in E. Sawicki, J. D. Mulik and E. Wittgenstein (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 1, Ann Arbor Sci. Publ., Ann Arbor, MI, 1978, p. 141.
246 K. Fujimura and M. Tsuchiya, *Bunseki Kagaku*, 37 (1988) 59.
247 L. J. Holcombe, B. F. Jones, E. E. Ellsworth and F. B. Meserole, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 401.
248 D. A. Stern, B. M. Myatt, J. F. Lachowski and K. T. McGregor, *EPA 600/9-84/015*, U.S., Environmental Protection Agency, Cincinnati, 1984, p. 33.
249 *Application Note*, 12, Dionex, Sunnyvale, CA, 1978.
250 L. C. Speitel, J. C. Spurgeon and R. A. Filipczak, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 75.
251 W. Coerdt and E. Mainka, *Fresenius' Z. Anal. Chem.*, 320 (1985) 503.
252 H. Matusiewicz and D. F. S. Natusch, *Int. J. Environ. Anal. Chem.*, 8 (1980) 227.
253 *Application Note*, 15, Dionex, Sunnyvale, CA, 1979.
254 D. Noel, H. Roberge and J. Hechler, *Anal. Chim. Acta*, 217 (1989) 135.
255 F. E. Butler, F. J. Toth, D. J. Driscoll, J. N. Hein and R. H. Jungers, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 185.
256 C. S. Mizisin, D. E. Kuivinen and D. A. Otterson, in J. D. Mulik and E. Sawicki (Editors), *Ion Chromatographic Analysis of Environmental Pollutants*, Vol. 2, Ann Arbor Sci. Publ., Ann Arbor, MI, 1979, p. 129.
257 P. Viswanadham, D. R. Smick, J. J. Pisney and W. Dilworth, *Anal. Chem.*, 54 (1982) 2431.
258 M. J. McCormick, *Anal. Chim. Acta*, 121 (1980) 233.
259 V. Abraham and J. M. deMan, *J. Am. Oil Chem. Soc.*, 64 (1987) 384.
260 V. B. Conrad and W. D. Brownlee, *Anal. Chem.*, 60 (1988) 365.
261 J. A. Cox and R. Saari, *Analyst (London)*, 112 (1987) 321.
262 H. Honma, K. Suzuki, M. Yoshida and H. Yanashima, *Bunseki Kagaku*, 37 (1988) T55.
263 D. W. Hausler and L. T. Taylor, *Anal. Chem.*, 53 (1981) 1227.
264 K. I. Burns, S. Elchuck and D. W. Everall, *First Ion Chromatography Forum, Boston, MA, Sept. 25, 1988*, Abstract No. 43.